

WEEK 1: Morning (9-12)	Monday	Tuesday	Wednesday	Thursday	Friday
	<p>Course intro</p> <p><i>Short talk (9-9.45):</i> "Welcome and introduction to course"</p> <p><i>"Speed-networking"(10-11):</i> "Get to know your fellow course participants"</p> <p><i>Lecture (11-12):</i> "Intro to bash and the unix universe"</p>	<p>Getting started with Python</p> <p><i>Short talk (9-9.30):</i> "Python introduction"</p> <p><i>Practical (9.30-10.30):</i> "Python introduction"</p> <p><i>Lecture (10.30-10.50):</i> "Python as a bioinformatical tool"</p> <p><i>Practical (10.50-12):</i> "Getting started with python"</p>	<p>Sequencing and raw-data QC</p> <p><i>Lecture (9.00 - 9.30):</i> "Introduction to sequencing"</p> <p><i>Lecture (9.40 – 10.45):</i> "Overview of sequencing technologies"</p> <p><i>Interactive Lecture/Practical (11.00 – 12.00):</i> "Sequencing Quality"</p>	<p>Genome assembly</p> <p><i>Lecture (9-9.30):</i> "Genome assembly strategies"</p> <p><i>Practical (9.30 -11):</i> "Bacterial genome assembly with short reads"</p> <p><i>Practical (11-12):</i> "Basic assembly QC stats"</p>	<p>Genome assembly</p> <p><i>Lecture (9-9.30):</i> "Intro to long-read and hybrid genome assembly"</p> <p><i>Practical (9.30-11):</i> "Long-read and hybrid assembly"</p> <p><i>Practical (11-12):</i> "Genome assembly comparison"</p>

WEEK 1: Afternoon (13-16.30)	Monday	Tuesday	Wednesday	Thursday	Friday
	<p>Getting started with the command-line and Bash</p> <p><i>Short talk (13-13.20):</i> Command-line introduction</p> <p><i>Practical (13.20-14.20):</i> Getting started with the command-line</p> <p><i>Short talk (14.30-14.45):</i> Bash introduction</p> <p><i>Practical (14.45-16.30):</i> Getting started with bash</p>	<p>Advancing in programming</p> <p><i>Lecture (13-13.20):</i> “Programming recap and Biopython deep-dive”</p> <p><i>Coding session (13.20-16.20):</i> “Multiple exercises command-line, bash and python” (participants choice)</p> <p><i>Lecture (16.20-16.30):</i> “Programming wrap-up”</p>	<p>Sequencing and raw-data QC</p> <p><i>Lecture/Practical (13.00-15.00):</i> “Quality control of sequencing data”</p> <p><i>Lecture/Practical (15:00-16.30):</i> “Contamination control, sample aggregation, and wrap-up”</p>	<p>Genome assembly</p> <p><i>Lecture/practical (13-14.00):</i> “Contamination”</p> <p><i>Coding session (14.00-16.30):</i> Biopython for assembly analysis</p>	<p>Genome analysis intro</p> <p><i>Lecture/Practical (13-15):</i> “Blast, in-silico MLST and gene annotation”</p> <p><i>Coding session (15-16.30):</i> Biopython continued, or practical catch-up (Participants choice)</p>

WEEK 2: Morning	Monday	Tuesday	Wednesday	Thursday	Friday
	<p>AMR</p> <p><i>Lecture (9-10):</i> “WGS-based detection of antimicrobial resistance (AMR) in bacteria”</p> <p><i>Practical (10.20-12):</i> “Use of bioinformatics tools and databases for WGS-based detection of AMR in bacteria”</p>	<p>Getting started with phylogenies</p> <p><i>Lecture (9-10):</i> “Evolution and phylogenies”</p> <p><i>Practical (10.20-12):</i> “Sequence handling and alignment”</p>	<p>Pipeline development</p> <p><i>Lecture (9-9.45):</i> “Basic principles of automation”</p> <p><i>Practical (10.00-11.45):</i> “Pipeline development – scaling number of tools and samples”</p>	<p>Data sharing</p> <p><i>Lecture (9-9.45):</i> “Introduction to sequence databases and data sharing”</p> <p><i>Practical (10.00-11.15):</i> “Uploading raw sequence data to ENA”</p> <p><i>Practical (11.15-12):</i> “Data exploration and retrieval using ENA resources”</p>	<p>Whats next?</p> <p><i>Lecture (9-9.45):</i> “Outlook on genome sequencing for disease surveillance”</p> <p><i>Site-visits (10-11):</i> Tour of the sequencing labs at SSI, meet the staff</p> <p><i>Mentor-chats (11-12):</i> Individual chats with participants to discuss continued learning. Free code session for remaining participants.</p>

WEEK 2: Afternoon	Monday	Tuesday	Wednesday	Thursday	Friday
	Tuberculosis profiling <i>Lecture (13-14):</i> “Tuberculosis - the world's deadliest curable disease” <i>Practical (14-16.30):</i> “ <i>M.tuberculosis</i> outbreak investigation and antibiotic resistance determination with TB-profiler”	Getting started with phylogenies <i>Lecture (13-14):</i> “Phylogenetic methods” <i>Practical (14.30-16.30):</i> “Inferring phylogenies”	Pipeline development <i>Practical (12.45-15.30):</i> “Pipeline development – from raw reads to data sharing” <i>Coding session (15.30-16.30):</i> Further exercises on writing pipelines (participants choice)	Data sharing <i>Lecture (13-13.45):</i> “Intro to online tools for genome analysis, data sharing and visualization, developed by ECDC) <i>Practical (14-16.30):</i> “Exploring Microreact, EpiPulse and Tessy”	Whats next? <i>Mentor-chats (13-14):</i> Individual chats with participants to discuss continued learning. Free code session for remaining participants. <i>Evaluation survey (14-14.30):</i> “Course evaluation” <i>Mingle and wrap-up</i>