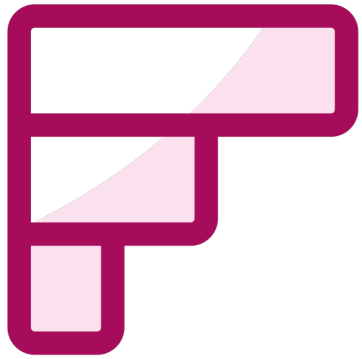


Additional Analysis

cgMLST (Core Genome MLST)



**What best describes your
experience with bioinformatics or
computational tools?**



Did you run cgMLST-based typing?



Do the cgMLST results support or contradict the SNP-based tree analysis?

cgMLST: Results

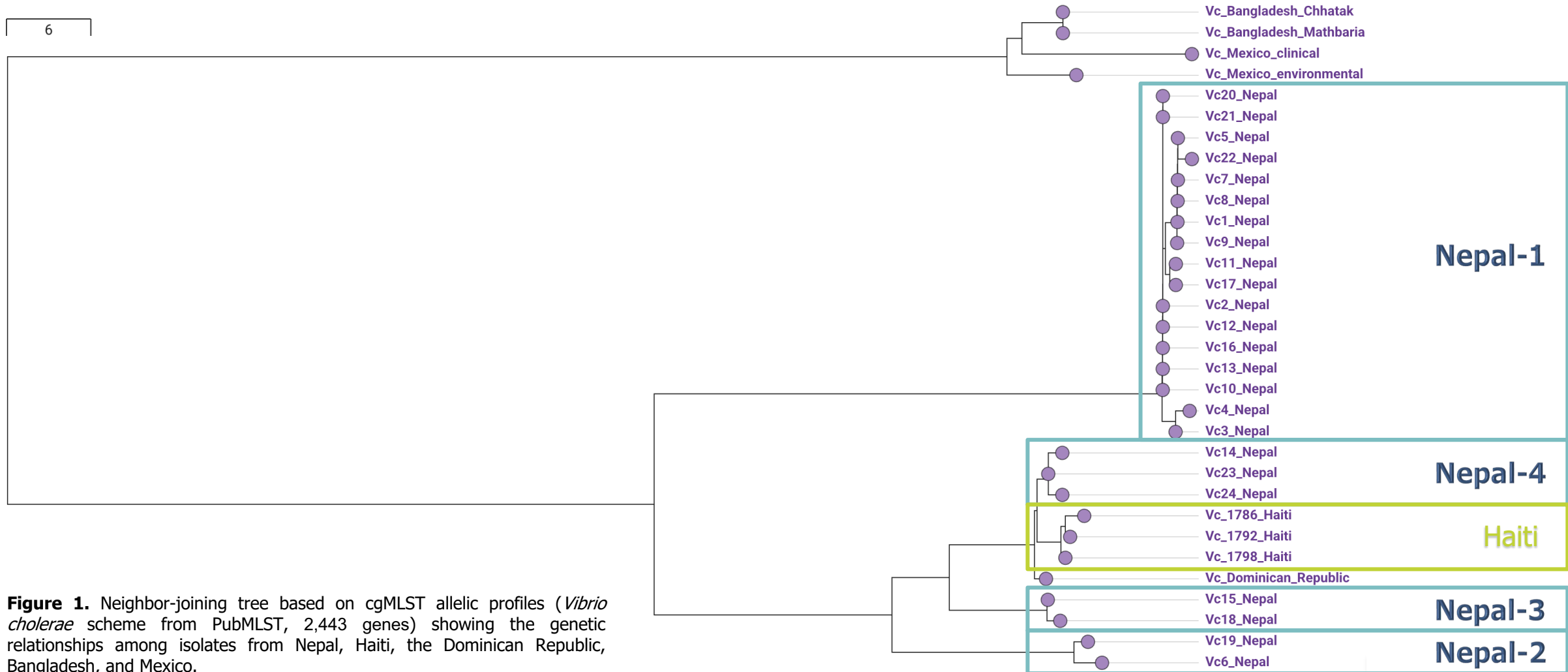


Figure 1. Neighbor-joining tree based on cgMLST allelic profiles (*Vibrio cholerae* scheme from PubMLST, 2,443 genes) showing the genetic relationships among isolates from Nepal, Haiti, the Dominican Republic, Bangladesh, and Mexico.

cgMLST: Results

Name	Vc10_N epal	Vc11_N epal	Vc12_N epal	Vc13_N epal	Vc14_N epal	Vc15_N epal	Vc16_N epal	Vc17_N epal	Vc18_N epal	Vc19_N epal	Vc1_N epal	Vc20_N epal	Vc21_N epal	Vc22_N epal	Vc23_N epal	Vc24_N epal	Vc2_N epal	Vc3_N epal	Vc4_N epal	Vc5_N epal	Vc6_N epal	Vc7_N epal	Vc8_N epal	Vc9_N epal	Vc_1786 Haiti	Vc_1792 Haiti	Vc_1798 Haiti
Vc10_Nepal	0	1	0	0	65	64	0	0	65	67	1	0	0	2	64	65	0	1	2	1	68	1	1	1	67	65	65
Vc11_Nepal	1	0	1	1	66	65	1	0	66	68	1	1	1	3	65	66	1	2	3	2	69	2	2	1	68	66	66
Vc12_Nepal	0	1	0	0	65	64	0	0	65	67	1	0	0	2	64	65	0	1	2	1	68	1	1	1	67	65	65
Vc13_Nepal	0	1	0	0	65	64	0	0	65	67	1	0	0	2	64	65	0	1	2	1	68	1	1	1	67	65	65
Vc14_Nepal	65	66	65	65	0	15	65	65	16	26	66	66	66	68	1	2	65	66	67	66	27	66	66	66	5	4	3
Vc15_Nepal	64	65	64	64	15	0	64	64	1	25	65	65	65	67	14	15	64	65	66	65	26	65	65	65	16	15	14
Vc16_Nepal	0	1	0	0	65	64	0	0	65	67	1	0	0	2	64	65	0	1	2	1	68	1	1	1	67	65	65
Vc17_Nepal	0	0	0	0	65	64	0	0	65	67	1	0	0	2	64	65	0	1	2	1	68	1	1	1	67	65	65
Vc18_Nepal	65	66	65	65	16	1	65	65	0	26	66	65	65	67	15	16	65	66	67	66	27	66	66	66	17	16	15
Vc19_Nepal	67	68	67	67	26	25	67	67	26	0	68	67	67	69	25	26	67	68	69	68	3	68	68	68	27	26	25
Vc1_Nepal	1	1	1	1	66	65	1	1	66	68	0	1	1	1	65	66	1	2	3	0	69	0	0	0	68	66	66
Vc20_Nepal	0	1	0	0	66	65	0	0	65	67	1	0	0	2	65	66	0	1	2	1	68	1	1	1	67	65	65
Vc21_Nepal	0	1	0	0	66	65	0	0	65	67	1	0	0	2	65	66	0	1	2	1	68	1	1	1	67	65	65
Vc22_Nepal	2	3	2	2	68	67	2	2	67	69	1	2	2	0	67	68	2	3	4	1	70	1	1	1	69	67	67
Vc23_Nepal	64	65	64	64	1	14	64	64	15	25	65	65	65	67	0	1	64	65	66	65	26	65	65	65	4	3	2
Vc24_Nepal	65	66	65	65	2	15	65	65	16	26	66	66	66	68	1	0	65	66	67	66	27	66	66	66	5	4	3
Vc2_Nepal	0	1	0	0	65	64	0	0	65	67	1	0	0	2	64	65	0	1	2	1	68	1	1	1	67	65	65
Vc3_Nepal	1	2	1	1	66	65	1	1	66	68	2	1	1	3	65	66	1	0	1	2	69	2	2	2	68	66	66
Vc4_Nepal	2	3	2	2	67	66	2	2	67	69	3	2	2	4	66	67	2	1	0	3	70	3	3	3	69	67	67
Vc5_Nepal	1	2	1	1	66	65	1	1	66	68	0	1	1	1	65	66	1	2	3	0	69	0	0	0	68	66	66
Vc6_Nepal	68	69	68	68	27	26	68	68	27	3	69	68	68	70	26	27	68	69	70	69	0	69	69	69	28	27	26
Vc7_Nepal	1	2	1	1	66	65	1	1	66	68	0	1	1	1	65	66	1	2	3	0	69	0	0	0	68	66	66
Vc8_Nepal	1	2	1	1	66	65	1	1	66	68	0	1	1	1	65	66	1	2	3	0	69	0	0	0	68	66	66
Vc9_Nepal	1	1	1	1	66	65	1	1	66	68	0	1	1	1	65	66	1	2	3	0	69	0	0	0	68	66	66
Vc_1786_Haiti	67	68	67	67	5	16	67	67	17	27	68	67	67	69	4	5	67	68	69	68	28	68	68	68	0	1	1
Vc_1792_Haiti	65	66	65	65	4	15	65	65	16	26	66	65	65	67	3	4	65	66	67	66	27	66	66	66	1	0	0
Vc_1798_Haiti	65	66	65	65	3	14	65	65	15	25	66	65	65	67	2	3	65	66	67	66	26	66	66	66	1	0	0

Table 1. Score Matrix associated with the cgMLST-based Neighbor-Joining (NJ) tree. It contains scaled pairwise genetic distances between genomes, calculated from substitution differences across the 2443 core loci, and adjusted to the expected core genome size to account for assembly completeness. Lower numbers indicate closer relationships.



Genomic Tools in Outbreak Investigations - Case Study Cholera Outbreak

Annotation: AMR genes, Virulence factors, and Genetic Elements

September 03, 2025

Intended Learning Objectives

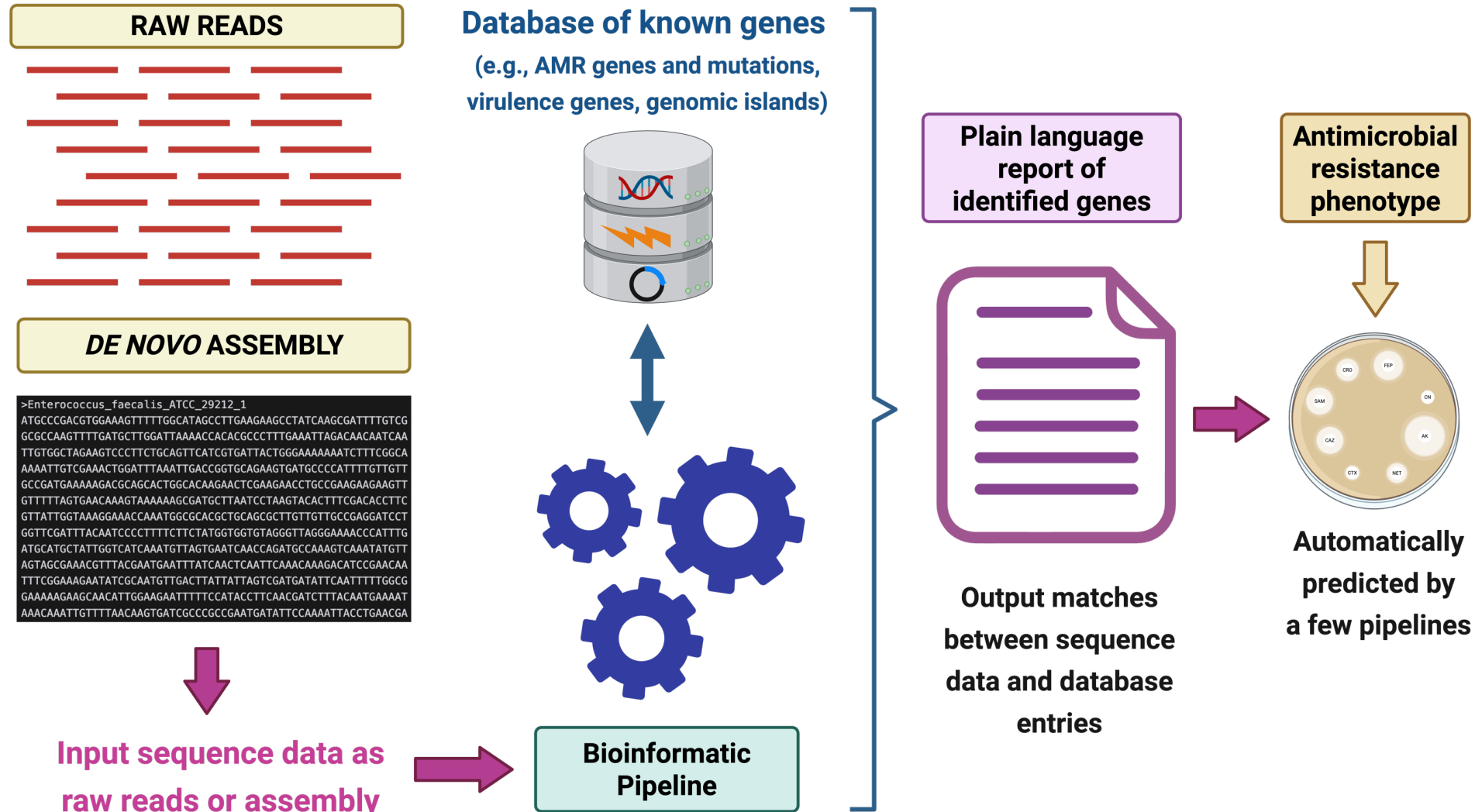
Specific objectives of this session:

1. Predict and interpret AMR and virulence genes from bacterial genomes.
2. Predict genomic islands (GIs) using IslandViewer4 and understand the limits of automated predictions.
3. Detect integrative conjugative elements (ICEs) using ICEfinder and interpret the main result views.
4. Understand that sequence alignment and literature comparison may be needed to identify known genetic elements in *Vibrio cholerae*.

This session consists of the following elements:

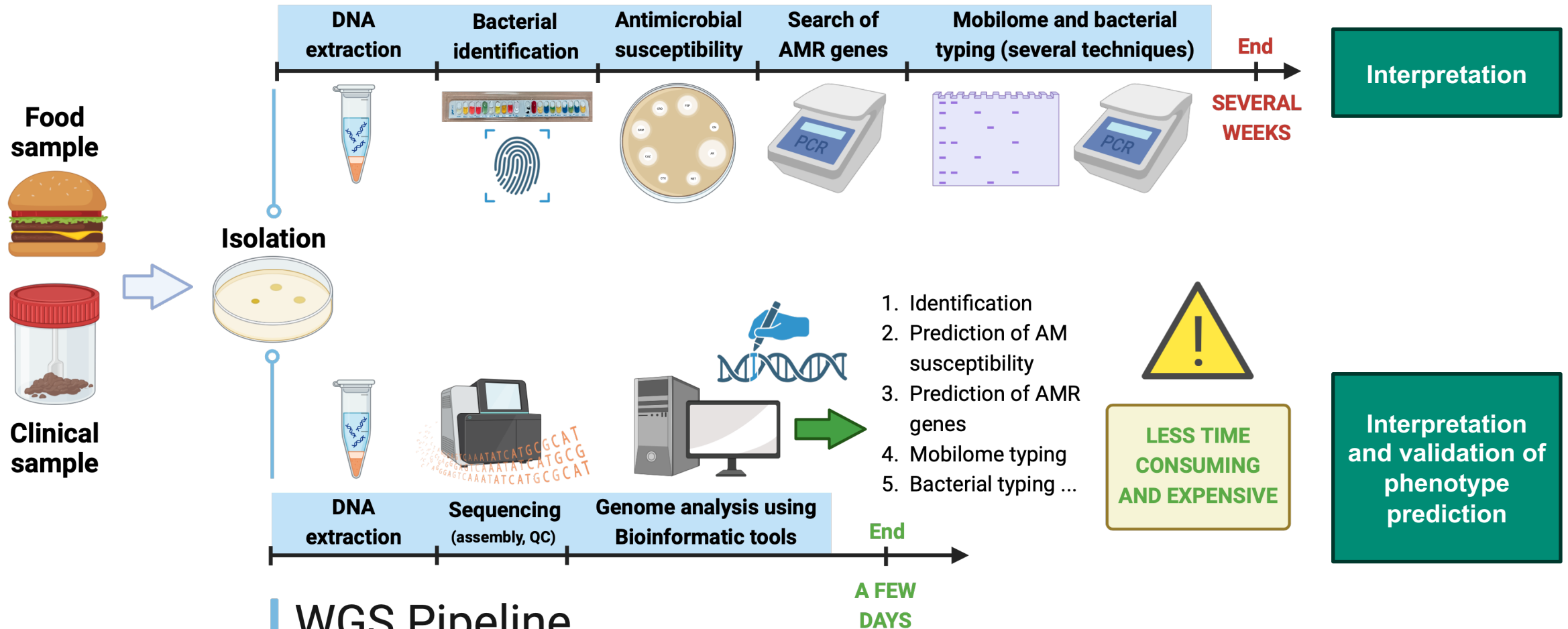
1. Introduction to genome annotation for AMR and virulence genes
2. Predicting ICEs using ICEfinder: input, outputs, and interpretation
3. Predicting genomic islands (GIs) with IslandViewer4: maps, tables, and region details
4. Understanding the limits of automated predictions and the need for literature and sequence alignment to identify known GI families (e.g., in *Vibrio cholerae*)

In silico Bacterial Genome Annotation



Why using WGS for Genome Analysis?

Traditional Pipeline



Annotation

Detection of Antimicrobial Resistance (AMR) genes and Virulence factors



Which AMR genes detection tool(s) did you run?



What identity and coverage thresholds did you apply for AMR gene detection?

AMR Phenotype/Genotype Prediction Tools

Table. Most used (command line-C or server-based-SB) AMR prediction tools for bacterial genomes

Tool name	Target	Input format	Link
ResFinder (C, SB)	General AMR (genotype, phenotype)	FASTA, FASTQ	http://genepi.food.dtu.dk/resfinder
PointFinder (C, SB)	Selected mutations (genotype)	FASTA, FASTQ	http://genepi.food.dtu.dk/resfinder
AMRFinderPlus (C)	General AMR (genotype) + <u>virulence</u>	FASTA	https://github.com/ncbi/amr/wiki
RGI (CARD) (C, SB)	General AMR (genotype)	FASTA	https://card.mcmaster.ca/
ABRicate (C)	Multiple databases (AMR, virulence, plasmids)	FASTA	https://github.com/tseemann/abricate
BV-BRC (SB, uses RAST)	General Annotation (genotype, phenotype)	FASTA	https://www.bv-brc.org/
Pathogenwatch (SB)	General AMR (genotype, phenotype) + <u>virulence</u>	FASTA, FASTQ	https://pathogen.watch/



Center for Genomic Epidemiology

Home

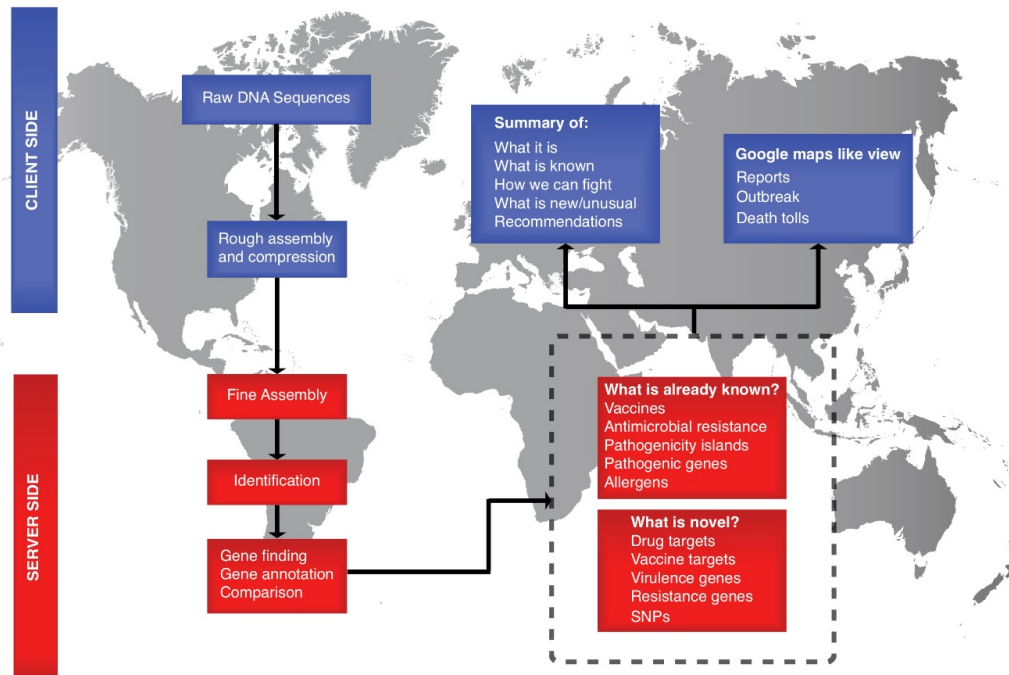
Services

Publications

Contact

Click **SERVICES** to see all the available tools

1



News

Scaling neighbor joining to one million taxa with dynamic and heuristic neighbor joining

January 2023

[Link to article....](#)

SourceFinder: a Machine-Learning-Based Tool for Identification of Chromosomal, Plasmid, and Bacteriophage Sequences from Assemblies

November 2022

[Link to article....](#)

PlasmidHostFinder: prediction of plasmid hosts using random forest

April 2022

[Link to article....](#)

ResFinder – an open online resource for identification of antimicrobial resistance genes in next-generation sequencing data and prediction of phenotypes from genotypes

January 2022

Phenotyping

2

[ResFinder](#)

Identification of acquired antibiotic resistance genes, and chromosomal gene mutations

[ResFinderFG](#)

Identification of functional metagenomic antibiotic resistance determinants.

[LRE-finder](#)

Identification of genes and mutations leading to linezolid resistance.

[KmerResistance](#)

Identification of acquired antibiotic resistance genes using Kmers.

ResFinder: Job Upload

ResFinder

Version

4.5.0 

ResFinder identifies acquired genes and/or finds chromosomal mutations mediating antimicrobial resistance in total or partial DNA sequence of bacteria.

ResFinder software: (2024-03-22)

ResFinder database: (2024-03-22)

PointFinder database: (2024-03-08)

DisinFinder database: (2023-05-31)

Chromosomal point mutations:

Threshold for %ID

90% 

Minimum length

60% 



Show unknown mutations



Ignore premature stop codons:



Ignore frameshift indels:

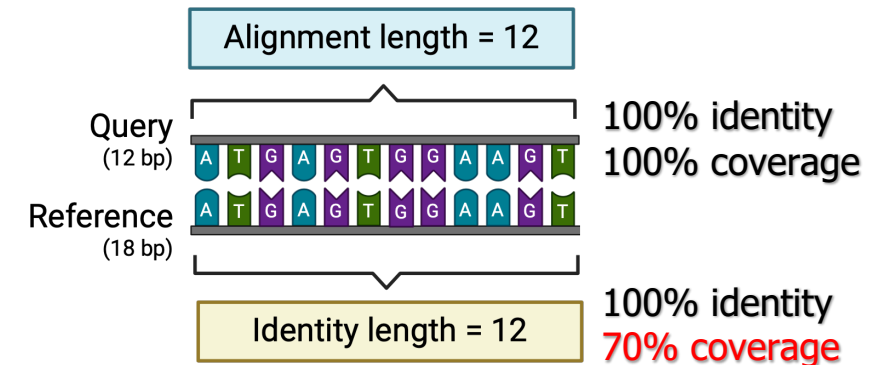
Acquired antimicrobial resistance genes:

Threshold for %ID

90% 

Minimum length

60% 



3

% **identity** and **coverage** can be adjusted if necessary

Click to view **known** and **unknown** mutations associated with AMR

ResFinder: Job Upload

Species and input data type:

Select species

Other

Select input type

FASTA (Assembled Genome/Contigs)

✓ FASTA (Assembled Genome/Contigs)

FASTQ (Non-nanopore Reads)

FASTQ (Nanopore Reads)

Disinfectant:

☐ Run disinfectant

Threshold for %ID

90%

Minimum length

60%

Upload and submit job:

Email (Get email, when finished - Optional):

Enter your email address...

Files (The sum of uploaded file sizes cannot exceed 1 gb):

Choose file No file chosen

Choose file No file chosen

Submit Job

*ResFinder phenotype predictions were validated using MIC values (BMD or Etest) and WGS data (Illumina sequencing). Phenotypic AST results were mainly interpreted using the EUCAST epidemiological cut-off values (ECOFFs) to categorize isolates as **WT** (MIC≤ECOFF) and **non-WT** (MIC>ECOFF).

Species-relevant prediction of **clinically relevant phenotypes*** for selected antimicrobial agents

Species and input data type:

Select species

- ✓ Other
- Campylobacter jejuni*
- Campylobacter coli*
- Campylobacter spp.*
- Enterococcus faecalis*
- Enterococcus faecium*
- Escherichia coli*
- Helicobacter pylori*
- Klebsiella*
- Mycobacterium tuberculosis*
- Neisseria gonorrhoeae*
- Plasmodium falciparum*
- Salmonella spp.*
- Staphylococcus aureus*

ResFinder: Job Upload

Species and input data type:

Select species

Other ▼

Select input type

FASTA (Assembled Genome/Contigs) ▼

Disinfectant:

☒ Run disinfectant

Threshold for %ID

90% ▼

Minimum length

60% ▼

Upload and submit job:

Email (Get email, when finished - Optional):

Enter your email address...

Files (The sum of uploaded file sizes cannot exceed 1 gb):

Choose file No file chosen

Choose file No file chosen

Submit Job

6

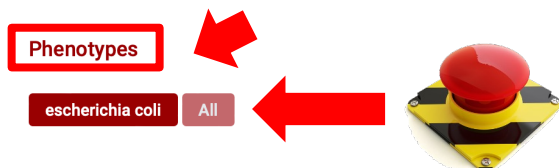
Write your **email** to receive the results when finished

7

Upload the **.fasta** or **.fastq** files of your bacterial genome for analysis

8

ResFinder: Analysis of Results



Hide

Antimicrobial	Class	WGS-predicted phenotype	Genetic background
gentamicin	aminoglycoside	No resistance	blaTEM-1A;;1;HM749966
tobramycin	aminoglycoside	No resistance	
amikacin	aminoglycoside	No resistance	
ciprofloxacin	quinolone	Resistant	
nalidixic acid	quinolone	Resistant	
ampicillin	beta-lactam	Resistant	
ampicillin+clavulanic acid	beta-lactam	No resistance	
cefepime	beta-lactam	No resistance	
cefotaxime	beta-lactam	No resistance	
cefoxitin	beta-lactam	No resistance	
ceftazidime	beta-lactam	No resistance	sul1;;2;U12338
ertapenem	beta-lactam	No resistance	
imipenem	beta-lactam	No resistance	
meropenem	beta-lactam	No resistance	
piperacillin+tazobactam	beta-lactam	No resistance	
temocillin	beta-lactam	No resistance	
sulfamethoxazole	folate pathway antagonist	Resistant	
trimethoprim	folate pathway antagonist	Resistant	
fosfomycin	fosfomycin	No resistance	dfrA1;;10;AF203818
azithromycin	macrolide	No resistance	
tetracycline	tetracycline	Resistant	tet(M);;5;U58985
tigecycline	tetracycline	No resistance	
chloramphenicol	amphenicol	Resistant	cmIA1;;1;M64556
colistin	polymyxin	Resistant	

Prediction of species-specific *in silico* antibiograms



Only **known** AMR mechanisms are detected



When using ("**All**" option) interpretation of results must be executed carefully → knowledge on **intrinsic resistance** (often mediated by structural traits) is **essential**

ResFinder: Analysis of Results

Acquired AMR gene hits

Hide

Resistance gene	Identity	Alignment length/gene length	Position in reference	Contig or depth	Position in contig	Phenotype	PMID	Accession no.	Notes
aadA2b	99.87%	780 / 780	1...780	DTU_2022_1023502_1_SI_S21FP06270_NODE_159_length_4297_cov_16.349400	1726...2505	['streptomycin', 'spectinomycin']	85862590	D43625	
aph(3')-Ia	100.00%	816 / 816	1...816	DTU_2022_1023502_1_SI_S21FP06270_NODE_216_length_1293_cov_9.157804	214...1029	['kanamycin', 'neomycin', 'lividomycin', 'paromomycin', 'ribostamycin']	6270337	V00359	
aadA1	100.00%	722 / 792	1...722	DTU_2022_1023502_1_SI_S21FP06270_NODE_245_length_737_cov_42.177049	16...737	['streptomycin', 'spectinomycin']	23180481	JQ414041	Alternative name ant(3'')-Ia
blaTEM-1A	100.00%	861 / 861	1...861	DTU_2022_1023502_1_SI_S21FP06270_NODE_19_length_64335_cov_9.271337	4073...4933	['amoxicillin', 'ampicillin', 'piperacillin', 'ticarcillin', 'cephalothin']	21393132	HM749966	Class A
mcr-1.1	100.00%	1626 / 1626	1...1626	DTU_2022_1023502_1_SI_S21FP06270_NODE_179_length_2787_cov_8.350000	239...1864	['colistin']	26603172	KP347127	
cmlA1	99.92%	1260 / 1260	1...1260	DTU_2022_1023502_1_SI_S21FP06270_NODE_159_length_4297_cov_16.349400	205...1464	['chloramphenicol']	1648560	M64556	MFS transporter
sul1	100.00%	840 / 840	1...840	DTU_2022_1023502_1_SI_S21FP06270_NODE_144_length_6159_cov_10.065816	702...1541	['sulfamethoxazole']	unpublished	U12338	
sul3	100.00%	792 / 792	1...792	DTU_2022_1023502_1_SI_S21FP06270_NODE_155_length_4607_cov_16.789286	1938...2729	['sulfamethoxazole']	12604565	AJ459418	
tet(M)	99.17%	1920 / 1920	1...1920	DTU_2022_1023502_1_SI_S21FP06270_NODE_180_length_2631_cov_8.655351	522...2441	['tetracycline', 'doxycycline', 'minocycline']	9372426	U58985	

ResFinder: Analysis of Results

Acquired AMR gene hits

Resistance gene	Identity	Alignment length/gene length	Position in reference	Contig or depth
aadA2b	99.87%	780 / 780	1...780	DTU_2022_1023502_1_SI_S21FP06270_NODE_159_length_4297_cov_16.349400
aph(3')-Ia	100.00%	816 / 816	1...816	DTU_2022_1023502_1_SI_S21FP06270_NODE_216_length_1293_cov_9.157804
aadA1	100.00%	722 / 792	1...722	DTU_2022_1023502_1_SI_S21FP06270_NODE_245_length_737_cov_42.177049
blaTEM-1A	100.00%	861 / 861	1...861	DTU_2022_1023502_1_SI_S21FP06270_NODE_245_length_737_cov_9.271337
mcr-1.1	100.00%	1626 / 1626	1...1626	DTU_2022_1023502_1_SI_S21FP06270_NODE_179_length_270_cov_10.065816
cmlA1	99.92%	1260 / 1260	1...1260	DTU_2022_1023502_1_SI_S21FP06270_NODE_159_length_4297_cov_16.349400
sul1	100.00%	840 / 840	1...840	DTU_2022_1023502_1_SI_S21FP06270_NODE_144_length_6159_cov_10.065816
sul3	100.00%	792 / 792	1...792	DTU_2022_1023502_1_SI_S21FP06270_NODE_155_length_4607_cov_16.789286
tet(M)	99.17%	1920 / 1920	1...1920	DTU_2022_1023502_1_SI_S21FP06270_NODE_180_length_2631_cov_8.655351

The number of aligned bases between the gene detected in your genome and the reference gene/total length of the reference gene in the database

The dark green color

Indicates a **perfect match** for a give gene. **Identity** is **100%** and the sequence in the genome **covers the entire length** of the resistance gene in the database

ResFinder: Analysis of Results

Acquired AMR gene hits

Resistance gene	Identity	Alignment length/gene length	Position in reference	Contig or depth
aadA2b	99.87%	780 / 780	1...780	DTU_2022_1023502_1_SI_S21FP06270_NODE_159_length_4297_cov_16.349400
aph(3')-Ia	100.00%	816 / 816	1...816	DTU_2022_1023502_1_SI_S21FP06270_NODE_159_length_4297_cov_16.349400
aadA1	100.00%	722 / 792	1...722	DTU_2022_1023502_1_SI_S21FP06270_NODE_245_length_737_cov_42.177049
blaTEM-1A	100.00%	861 / 861	1...861	DTU_2022_1023502_1_SI_S21FP06270_NODE_19_length_64335_cov_9.271337
mcr-1.1	100.00%	1626 / 1626	1...1626	DTU_2022_1023502_1_SI_S21FP06270_NODE_179_length_2787_cov_8.350000
cmlA1	99.92%	1260 / 1260	1...1260	DTU_2022_1023502_1_SI_S21FP06270_NODE_159_length_4297_cov_16.349400
sul1	100.00%	840 / 840	1...840	DTU_2022_1023502_1_SI_S21FP06270_NODE_159_length_4297_cov_16.349400
sul3	100.00%	792 / 792	1...792	DTU_2022_1023502_1_SI_S21FP06270_NODE_159_length_4297_cov_16.349400
tet(M)	99.17%	1920 / 1920	1...1920	DTU_2022_1023502_1_SI_S21FP06270_NODE_180_length_2631_cov_8.655351

The grey color

Indicates a **warning** due to a non-perfect match, % ID is 100% → alignment length **is different** than resistance gene length

The light green color

Indicates a **warning** due to a non-perfect match, % ID < 100% → alignment length **is equal** to the resistance gene length

ResFinder: Analysis of Results

Resistance gene	Identity	Alignment length/gene length	Position in reference	Contig or depth
aadA2b	99.87%	780 /	1...780	DTU_2022_1023502_1_SI_S21FP06270_NODE_159_length_4297_cov_16.349400
aph(3')-Ia	100.00%	816 /	1...816	DTU_2022_1023502_1_SI_S21FP06270_NODE_216_length_1293_cov_9.157804
aadA1	100.00%	722 /	1...722	DTU_2022_1023502_1_SI_S21FP06270_NODE_245_length_737_cov_42.177049
blaTEM-1A	100.00%	861 /	1...861	DTU_2022_1023502_1_SI_S21FP06270_NODE_19_length_64335_cov_9.271337
mcr-1.1	100.00%	1626 /	1...1626	DTU_2022_1023502_1_SI_S21FP06270_NODE_179_length_2787_cov_8.350000
cmlA1	99.92%	1260 /	1...1260	DTU_2022_1023502_1_SI_S21FP06270_NODE_159_length_4297_cov_16.349400
sul1	100.00%	840 /	1...840	DTU_2022_1023502_1_SI_S21FP06270_NODE_144_length_6159_cov_10.065816
sul3	100.00%	792 /	1...792	DTU_2022_1023502_1_SI_S21FP06270_NODE_155_length_4607_cov_16.789286
tet(M)	99.17%	1920 /	1...1920	DTU_2022_1023502_1_SI_S21FP06270_NODE_180_length_2631_cov_8.655351

Gene Alignments

Show gene alignments

Click "**Show Gene Alignments**" to see the alignment between the query and the reference gene

aph(3')-Ia

Reference sequence: ATGTTTCATGCCGCCTGTTTTCTGCTCATTGGCACGTTTCGCAACCTGT
Query sequence: ATGTTTCATGCCGCCTGTTTTCTGCTCATTGGCACGTTTCGCAACCTGT

Reference sequence: TCTCATTGCGGACACCTTTTCCAGCCTCGTTTGAAAGTTTCATTGCCAG
Query sequence: TCTCATTGCGGACACCTTTTCCAGCCTCGTTTGAAAGTTTCATTGCCAG

Reference sequence: ACGGGACTCCTGCAATCGTCAAGGGATTGAAACCTATAGAAGACATTGCT
Query sequence: ACGGGACTCCTGCAATCGTCAAGGGATTGAAACCTATAGAAGACATTGCT

ResFinder: Analysis of Results

Chromosomal mutations mediating AMR

Hide

Gene / Region	Identity	Alignment Length / Gene Length	Position in reference	Config or Depth	Position in config	PMID	Accession no.	Notes
gyrA	99.51%	2628 / 2628	1..2628	DTU_2022_1023502_1_SI_S21FP06270_NODE_47_length_37275_cov_8.114380	23787..26414	No PMID	CP073768.1	No notes

Mutation	Nucleotide change	Amino acid change	Phenotype	PMID	Notes
p.S83L	tcg>ttg	s -> l	ciprofloxacin,nalidixic acid	8891148,2168148,12654733	undefined
p.D87N	gac>aac	d -> n	ciprofloxacin,nalidixic acid	12654733,22878251,1850972	undefined

Gene / Region	Identity	Alignment Length / Gene Length	Position in reference	Config or Depth	Position in config	PMID	Accession no.	Notes
parC	99.87%	2259 / 2259	1..2259	DTU_2022_1023502_1_SI_S21FP06270_NODE_51_length_35503_cov_8.962517	6191..8449	No PMID	CP084529.1	No notes

Mutation	Nucleotide change	Amino acid change	Phenotype	PMID	Notes
p.S80I	agc>atc	s -> i	ciprofloxacin,nalidixic acid	8851598,21856834-20638608,8524852,25631675	undefined

Gene / Region	Identity	Alignment Length / Gene Length	Position in reference	Config or Depth	Position in config	PMID	Accession no.	Notes
gyrB	99.13%	2415 / 2415	1..2415	DTU_2022_1023502_1_SI_S21FP06270_NODE_29_length_49656_cov_8.154071	33866..36280	No PMID	CP047010.1	No notes

Mutations known to confer resistance to antibiotics will appear below the corresponding gene (e.g., *gyrA* or *parC* mutations)

No known mutations detected

ResFinder: Analysis of Results

Downloads

Table downloads

Download phenotypetable (txt) Download species specific phenotype table (txt)

Download acquired AMR gene results:

Results as text Hit in genome sequences Resistance gene sequences Results as tabseperated file

Download Chromosomal point mutation results:

Results as tabseperated file Results as text file

Download **all** or **species-specific** phenotypic and genotypic data in a *.txt* file format

Download of acquired **AMR genes** (*.txt* file format) and **gene sequences** (*.fasta* file format)

Download of chromosomal **point mutations** in a *.txt* file format

AMRFinderPlus

Software (command line) and the **database** allow the identification of **acquired antimicrobial resistance** genes in bacterial protein and/or assembled nucleotide sequences as well as **known resistance-associated point mutations** for several taxa

```
# Install AMRFinderPlus and download the last database
$ mamba install ncbi-amrfinderplus
$ amrfinder -U

# See a list of all available organisms
$ amrfinder --list_organisms

# Run AMRFinderPlus in the the assembled genome
$ amrfinder -n file_dna.fasta -O Vibrio_cholerae -o AMRFinderPlus_results

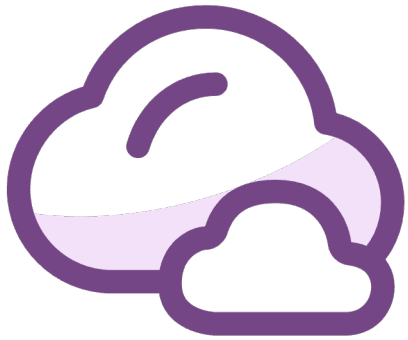
# Run AMRFinderPlus in the the assembled genome with the “plus” option (virulence and metal+biocide tolerance genes)
$ amrfinder -n file_dna.fasta -O Vibrio_cholerae --plus -o AMRFinderPlus_plus_results
```

AMRFinderPlus: Output

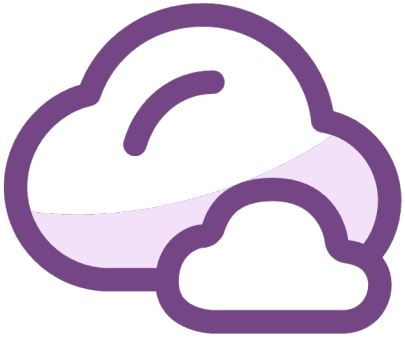
Contig id	Start	Stop	Strand	Element symbol	Element name	Scope	Type	Subtype	Class	Subclass	Method	Target length	Reference sequence length	% Coverage of reference	% Identity to reference	Alignment length	Closest reference accession	Closest reference name
NZ_LT90661 4.1	1566982	1567353	-	ctxB	cholera enterotoxin binding subunit CtxB	plus	VIRULENCE	VIRULENCE	NA	NA	EXACTX	124	124	100	100	124	AAF94613.1	cholera enterotoxin binding subunit CtxB
NZ_LT90661 4.1	1567353	1568126	-	ctxA	cholera enterotoxin catalytic subunit CtxA	plus	VIRULENCE	VIRULENCE	NA	NA	EXACTX	258	258	100	100	258	AAF94614.1	cholera enterotoxin catalytic subunit CtxA
NZ_LT90661 4.1	1674334	1675455	+	varG	VarG family subclass B1-like metallo-beta-lactamase	core	AMR	AMR	BETA-LACTAM	CARBAPENEM	EXACTX	374	374	100	100	374	WP_000778180.1	VarG family subclass B1-like metallo-beta-lactamase
NZ_LT90661 4.1	1687457	1688275	-	almG	glycine--lipid A transferase AlmG	core	AMR	AMR	COLISTIN	COLISTIN	EXACTX	273	273	100	100	273	WP_001881365.1	glycine--lipid A transferase AlmG
NZ_LT90661 4.1	1688280	1688513	-	almF	lipid A modification system glycine carrier protein AlmF	core	AMR	AMR	COLISTIN	COLISTIN	EXACTX	78	78	100	100	78	WP_000806242.1	lipid A modification system glycine carrier protein AlmF
NZ_LT90661 4.1	1688500	1690167	-	almE	lipid A modification system glycine--protein ligase AlmE	core	AMR	AMR	COLISTIN	COLISTIN	BLASTX	556	556	100	98.38	556	WP_252944138.1	lipid A modification system glycine--protein ligase AlmE
NZ_LT90661 5.1	315307	315933	+	catB9	type B-5 chloramphenicol O-acetyltransferase CatB9	core	AMR	AMR	PHENICOL	CHLORAMPHENICOL	EXACTX	209	209	100	100	209	WP_001009012.1	type B-5 chloramphenicol O-acetyltransferase CatB9



Did you observe similar AMR gene profiles in all isolates from Nepal?”



Which AMR genes can explain the phenotypic resistance profiles reported in the paper?”



Which do you think is more robust for detecting novel resistance variants: nucleotide-based, protein-based, both or none?”

Predicted AMR Genes and Mutations

Class	Antibiotics tested	Nepal isolates	AMR genes (Pathogenwatch)
β-lactams (penicillins & combinations)	Ampicillin		blaTEM (not present in all isolates), varG (intrinsic)
	Amoxicillin–clavulanic acid		varG (intrinsic)
β-lactams (cephalosporins, 3rd gen.)	Cefotaxime		varG (intrinsic)
	Ceftiofur		varG (intrinsic)
Fluoroquinolones	Ciprofloxacin	Decreased susceptibility	qnrVC1 (not present in all isolates), gyrA_S83I*, gyrA_S85L*
	Nalidixic acid	Resistant	gyrA_S83I*, gyrA_S85L*
Tetracyclines	Tetracycline		tet(59) (not present in all isolates)
Aminoglycosides	Apramycin		-
	Streptomycin		strA*, strB*
	Spectinomycin		-
	Gentamicin		-
	Neomycin		-
Phenicol	Chloramphenicol		catB9* (unexpressed), floR (not present in all isolates)
	Florfenicol		floR
Polymyxins	Colistin		-
Sulfonamides	Sulfamethoxazole	Resistant	sul2
Diaminopyrimidines	Trimethoprim	Resistant	dfrA1, dfrA31 (not present in all isolates)

*, Partial match to the gene in the database; <https://vibriowatch.readthedocs.io/en/latest/AMR.html>

CholeraeFinder

Center for Genomic Epidemiology

Username

Password

New Reset Login

Home

Services

Instructions

Output

Article abstract

CholeraeFinder 1.0

This tool is no longer being supported or maintained.

CholeraeFinder identifies Vibrio Cholerae serogroup/biotypes, and genes of interest for the pathologic types.

Select threshold for %ID

98 %

Select minimum length

Length a gene in the genome at least has to cover of the length of the gene in the database to be outputted

60 %

Select type of your reads

Assembled Genome/Contigs*

Isolate File

Name	Size	Progress	Status
------	------	----------	--------

Upload

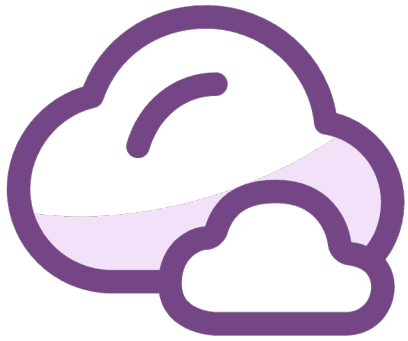
Remove

1

Upload your results in **FASTQ** or **FASTA** (assembled genomes) format. The results will take a few minutes to appear



Which virulence genes detection tool(s) did you run?



Which key virulence genes did you detect in the *V. cholerae* isolates from Nepal?



Did you notice any differences in virulence genes among the isolates in this study?

Virulence Genes: Results

Virulence gene (Pathogenwatch)	Phenotype	Notes
ace	Toxin	
acfA, acfB, acfC (~), acfD	Colonisation	acfC: Different gene sequence in isolates from Nepal-2, -3, -4, and Haiti
ctxA, ctxB	Toxin	
hapA	Haemagglutinin	
hlyA	Haemolysin	
makA	Toxin	
nanH	Neuraminidase	
ompU	Adhesion	
rpoS	Fitness (sigma factor)	
tagA (~)	Mucinase	Different gene sequence in isolates from Nepal-1
toxR	Regulator	
vasX	Toxin	
zot	Toxin	
TCP cluster	Colonisation	
Lux operon	Quorum sensing	
RTX Toxin Operon (~)	Toxin	Different gene sequence in isolates from Haiti
MSHA Pilus	Adhesion	

Annotation

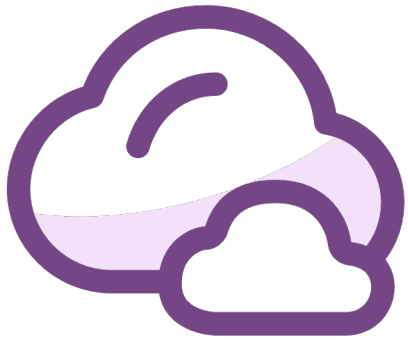
Identification of Genetic Elements



Were you able to detect any integrative conjugative elements (ICEs) or genomic islands in the isolates?



Which integrative conjugative element (ICE) family is typically found in 7th pandemic V. cholerae strains?



Which AMR genes did you find within the detected ICE?

ICEberg 3.0

Provide a comprehensive, curated database and analysis platform for **Integrative and Conjugative Elements (ICEs)** and related mobile genetic elements, allowing users to explore, identify, and compare ICEs across bacterial genomes



ICEberg 3.0

Go to old version ICEberg 2.0

MAIN NAVIGATION

- Welcome
- ICE Browse**
- mICE Browse
- Statistics
- Tools**
- Download
- Reference
- Help manual

Welcome to ICEberg 3.0!

Integrative and conjugative elements in bacteria

ICEberg 3.0: functional categorization and analysis of the integrative and conjugative elements in bacteria

Integrative and conjugative elements (ICEs) are essential mobile genetic elements that play a significant role in bacterial evolution. They can integrate into the bacterial chromosome and possess a complete conjugation machinery, enabling self-transmission between bacterial cells. By facilitating the horizontal transfer of diverse cargo genes, ICEs confer host beneficial traits such as antibiotic resistance, pathogenesis, defense system, metal resistance, compound degradation, and symbiosis, which contribute significantly to bacterial diversity and adaptation. Here, we report the release of ICEberg 3.0, which offers three major enhancements: (i) New ICE, IME and CIME data with manual curations. (ii) Categorization of ICE cargo functions. (iii) Extends its focus to encompass ICEs from the human microbiome. These updates enable efficient in facilitating the understanding of bacterial evolution by capturing the diverse functions and characteristics of ICEs.

Antibiotic resistance
Virulence factor
Metal resistance
Defense system
Degradation
Symbiosis

ICEs
IMEs CIMEs
mICEs

ICE Prediction

Functional annotation

News

- ICEberg 3.0 database is online. (Jun 1, 2023)
- mICE dataset update. (Feb 15, 2023)
- ICEfinder update. (Oct 15, 2022)
- Website restructuring. (Aug 28, 2022)

Citation

M. Wang, G Liu, M Liu, C Tai, Z. Deng, J Song, H.Y. Ou.
ICEberg 3.0: functional categorization and analysis of the integrative and conjugative elements in bacteria. *Nucleic Acids Research*.2024 Jan 5; 52(D1):D732-D737. doi: 10.1093/nar/gkad935

Contact

Microbial Bioinformatics Group, State Key Laboratory of Microbial Metabolism, Shanghai Jiao Tong University, 200030, China
Email: hyou@sjtu.edu.cn

ICEberg 3.0: Job Upload

Click on **Tools** → **ICEfinder** in the top menu



ICEberg 3.0 Go to old version ICEberg 2.0

MAIN NAVIGATION

- Welcome
- ICE Browse
- mICE Browse
- Statistics
- Tools**
 - ICEfinder**
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- Help manual

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Antibiotic resistance
Virulence factor
Metal resistance
Defense system
Degradation
Symbiosis

ICEs
IMEs CIMEs
mICEs

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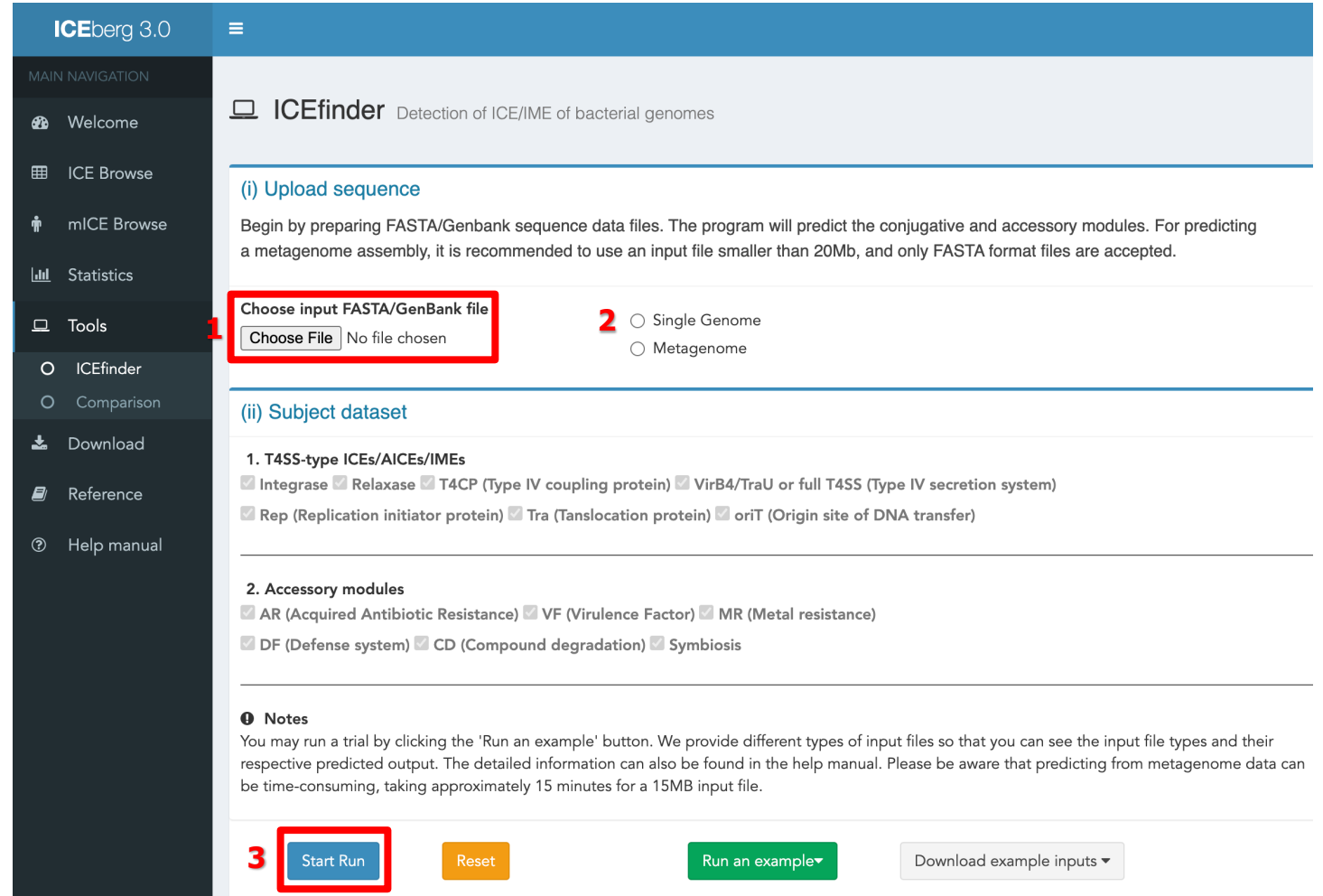
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Contact

Microbial Bioinformatics Group, State Key Laboratory of Microbial Metabolism, Shanghai Jiao Tong University, 200030, China
Email: hyou@sjtu.edu.cn

ICEberg 3.0: Job Upload

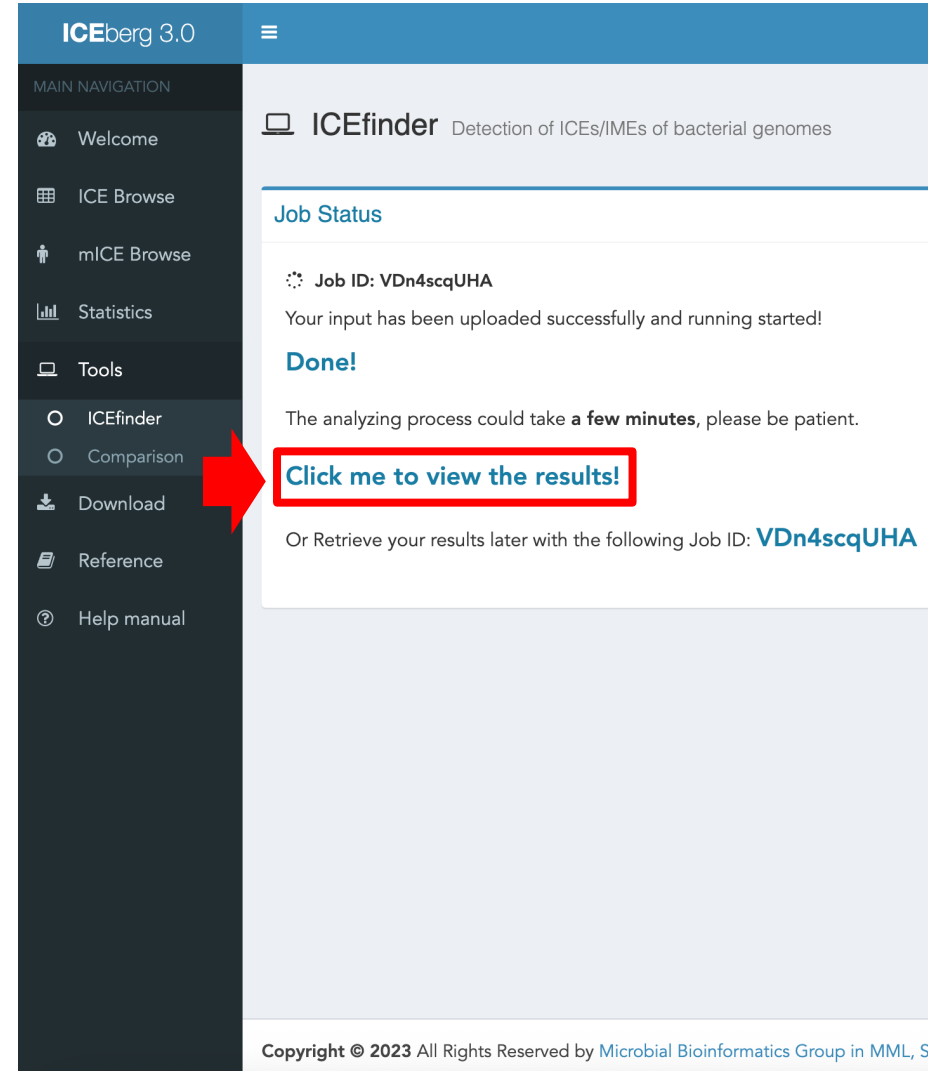
1. In ICEfinder, click “**Choose File**” and select the file containing your genome
2. Accepted formats include:
 - **GenBank** format (.gb, .gbk, .gbff) → recommended, since it contains both sequence and feature annotations
 - **FASTA** format (.fna, .fa) → accepted, but with fewer annotations (results may be less informative)
3. Click “**Single Genome**” (single cell finished genomes)
4. Press “**Start Run**”



The screenshot shows the ICEfinder web interface. On the left is a dark sidebar with 'MAIN NAVIGATION' links: Welcome, ICE Browse, mICE Browse, Statistics, Tools (highlighted with a red '1'), ICEfinder, Comparison, Download, Reference, and Help manual. The main content area has a blue header 'ICEfinder' with the subtitle 'Detection of ICE/IME of bacterial genomes'. Below this, section '(i) Upload sequence' contains instructions and a 'Choose input FASTA/GenBank file' section with a 'Choose File' button (highlighted with a red box and a red '2') and radio buttons for 'Single Genome' (selected) and 'Metagenome'. Section '(ii) Subject dataset' has two parts: '1. T4SS-type ICEs/AICEs/IMEs' with checkboxes for Integrase, Relaxase, T4CP, VirB4/TraU, Rep, Tra, and oriT; and '2. Accessory modules' with checkboxes for AR, VF, MR, DF, CD, and Symbiosis. A 'Notes' section at the bottom explains the trial run. At the very bottom, a 'Start Run' button (highlighted with a red box and a red '3') is next to a 'Reset' button, followed by 'Run an example' and 'Download example inputs' buttons.

ICEberg 3.0: Analysis of Results

- After uploading a genome file (FASTA or GenBank), ICEfinder starts an analysis job
- Each run is assigned a unique Job ID (**e.g. VDn4scqUHA**)
- The analysis takes a few minutes, depending on genome size
- When complete, ICEfinder shows “**Done!**” and provides a link
- Click to **view the results!**
- Results can also be retrieved later using the Job ID



ICEberg 3.0

MAIN NAVIGATION

- Welcome
- ICE Browse
- mICE Browse
- Statistics
- Tools
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- Comparison
- Download
- Reference
- Help manual

ICEfinder Detection of ICEs/IMEs of bacterial genomes

Job Status

Job ID: VDn4scqUHA

Your input has been uploaded successfully and running started!

Done!

The analyzing process could take a few minutes, please be patient.

[Click me to view the results!](#)

Or Retrieve your results later with the following Job ID: **VDn4scqUHA**

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ICEberg 3.0: Analysis of Results

Circle (right): genome map; blue bar = predicted ICE location

Table "Result View" (left):

- Region 1 → coordinates (125–151 kb), length ~26 kb
- Type: T4SS-type ICE = core ICE signatures detected: integrase, relaxase, T4CP, T4SS

Meaning: Genome contains one ICE-like region

ICEberg 3.0 Go to old version ICEberg 2.0

MAIN NAVIGATION

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- Help manual

ICEfinder Result Home > ICEfinder

JobID	VDn4scqUHA
Submission date	Wed Aug 27 05:36:31 2025
Sequence name	Vibrio cholerae O1 str. 2010EL-1786 chromosome 1, complete sequence
Length	3031375 bp
GC Content	47.67 %

Result Map



CP003069.1
3031375 bp

#	Region	Coords	Length (bp)	Type	Detail
1	Region1	125439..151544	26106	T4SS-type ICE	

Click to see the annotated ICE

Copyright © 2023 All Rights Reserved by Microbial Bioinformatics Group in MML, SJTU. Version 3.0

ICEberg 3.0: Analysis of Results

- **Type:** T4SS-type ICE → detected conjugation machinery (Type IV Secretion System)
- **Location:** coordinates 125–151 kb in the chromosome
- **Length:** ~26 kb region
- **GC Content:** 48.3 %, close to host genome
- **Relaxase type:** MOBH → enzyme that initiates DNA transfer
- **Mating pair formation system:** typeF → conjugative pilus system

ICEfinder Result	
I. Summary	
Type	T4SS-type ICE
Location (nt)	125439..151544
Length (bp)	26106
GC Content (%)	48.28
oriT seq	-
DRs	-
Relaxase Type	MOBH
Mating pair formation systems	typeF
Close to tRNA	-

Meaning: ICEfinder predicts one ICE-like region, identified as a T4SS-type element with MOBH relaxase and typeF conjugation system

ICEberg 3.0: Analysis of Results

- **Top graph:** GC% variation across the predicted ICE (differences may indicate foreign DNA)
- **Colored blocks** (bottom): functional genes/modules detected. E.g.,:
 - **Green** = defense system
 - **Pink** = T4SS genes
 - **Brown** = relaxase

Meaning: The region has the typical modular structure of an ICE: conjugation genes + accessory/defense genes



ICEberg 3.0: Analysis of Results

- **Each row** = one gene in the predicted ICE region
- **Columns:**
 - **Coordinates/size** → position on the genome
 - **Product** → predicted protein function
 - **Feature** → role in the ICE (defense, relaxase, T4CP, T4SS, etc.)

Meaning: Region has conjugation machinery + defense genes → typical ICE structure

III. Gene List

Gene	Coordinates [+/-], size (bp)	Product	Feature
Vch1786_I0098	115524..119265 [+], 3742	conserved BREX,pglX1	Flank; Defense
Vch1786_I0099	119277..121581 [+], 2305	PglZ domain protein; BREX,pglZA	Flank; Defense
Vch1786_I0100	121619..123671 [+], 2053	ATP-dependent Lon protease; BREX,brxL	Flank; Defense
Vch1786_I0101	123697..124468 [+], 772	conserved hypothetical protein	Flank
Vch1786_I0102	124502..125318 [+], 817	conserved AbiJ,AbiJ	Flank; Defense
Vch1786_I0103	125439..127590 [+], 2152	conjugal transfer pilus assembly protein TraI, MOBH	Relaxase
Vch1786_I0104	127638..129459 [+], 1822	conjugal transfer pilus assembly protein TraD, t4cp2	T4CP
Vch1786_I0105	129468..130029 [+], 562	plasmid-related protein	
Vch1786_I0106	130015..130651 [+], 637	plasmid-related protein, G_tfc7	T4SS
Vch1786_I0107	130677..131265 [-], 589	conserved hypothetical protein	
Vch1786_I0108	131278..131518 [-], 241	plasmid-related protein	
Vch1786_I0109	131553..131835 [+], 283	conjugal transfer pilus assembly protein TraL, F_traL	T4SS
Vch1786_I0110	131831..132458 [+], 628	conjugal transfer pilus assembly protein TraE, F_traE	T4SS

ICEberg 3.0: Analysis of Results

Download section:

- DNA (region sequence) or Protein (all predicted proteins) can be saved as FASTA files

Comparison section:

- Predicted ICE region can be compared with ICEberg's curated database

Two methods available: Mash (fast, k-mer) or Multigeneblast (gene content/synteny)

IV. Download

	No. of sequences	Download
DNA	1	Fasta
Protein	24	Fasta

V. Comparison

ICE Region: 125439..151544, 26106 bp

Comparison Method:

☒ Mash ☐ Multigeneblast

[Run](#)

Notes

The predicted ICEs region can be submitted for comparison with the ICEberg database. Two comparison methods are provided, namely Mash and Multigeneblast, both using default parameters. The Mash distance threshold is set to 0.005, while the parameters for Multigeneblast are as follows: hitsperegene 250, minseqcov 25, minpercid 30, distancekb 10, and syntenyweight 0.5.

Key point:

ICEberg detects ICE-like regions but does not always identify the ICE family.

- 👉 Extra “detective work” is needed — a good next step is to run **Multigeneblast** for similarity to known ICEs present in ICEberg database (“ICE Browse”)

ICEberg 3.0: Analysis of Results

- **Rows** = known ICEs; **colored arrows** = conserved genes
- **Total score** = weighted similarity of the ICE clusters (coverage + gene order + similarity)
- **Cumulative BLAST bit score** = sum of all gene-to-gene matches (raw sequence similarity)
- Predicted ICE matches several *V. cholerae* ICEs (several belonging to SXT/R391 family)

Key point: ICEberg suggests related ICEs but may miss full length

👉 Confirm family/boundaries by aligning the full ICE (e.g., with progressiveMauve)





Which genomic island(s) did you find in your isolates?

IslandViewer 4



Predicts **genomic islands** (foreign DNA regions mostly acquired by horizontal transfer) using **four methods** (IslandPick-comparative genomics, IslandPath-DIMOB-mobility genes, SIGI-HMM-codon usage bias, Islander-integration near tRNA genes)

IslandViewer 4 | An integrated interface for computational identification and visualization of genomic islands

[BROWSE](#) [ABOUT](#) [GENOME UPLOAD](#) [DOWNLOAD](#) [RESOURCES](#) [CONTACT US](#) [FAQ](#) [ACKNOWLEDGEMENTS](#) [HTTP API](#) [LOGIN](#)

If you use IslandViewer for your research, please cite the most recent publication. Thank You!

Bertelli, C. et al. 2017.

"IslandViewer 4: Expanded prediction of genomic islands for larger-scale datasets"

Nucleic Acids Research. 2017 May 2.

doi: [10.1093/nar/gkx343](https://doi.org/10.1093/nar/gkx343)

If you would like to do more comparative analysis of genomes/genomic islands try our new tool

[IslandCompare](#)

ISLANDVIEWER

IslandViewer is a computational tool that integrates four different genomic island prediction methods: IslandPick, IslandPath-DIMOB, SIGI-HMM, and Islander. For more information about methods and analyses see [here](#) and in the [FAQs](#).

Try our **user management interface** available under LOGIN (optional), and submit [here](#) your custom genome to predict genomic islands using IslandPick, IslandPath-DIMOB, and SIGI-HMM.

Or browse the pre-computed genomic island predictions for publicly available genomes, including Islander predictions as well:

[Go to Genome](#) [Example](#), [Example \(incomplete genome\)](#)

News

2024-12-13 - Yesterday there was an unscheduled downtime but this has now been rectified. We apologize for any inconvenience and if you have any further problems please don't hesitate to contact us.

2024-09-06 - The most recent update of the IslandViewer reference database has completed after making significant back-end changes to our update process so the update could occur with minimal downtime. This is a major update, and IslandViewer now has pre-computed GI results for 34,184 genomes. Please contact us at islandpick-mail@sfu.ca if you have any questions.

IslandViewer 4: Job Upload

1. **File format:** Use GenBank (.gbk/.gbff) or EMBL file
2. **Choose file:** Click "Choose File" and select your genome file.
3. (Optional) Add a genome name or email to be notified when the run finishes
4. Click "**Upload**"

Note: Whole genomes work best — fragmented assemblies may give false positives or miss GIs

IslandViewer 4 | An integrated interface for computational identification and visualization of genomic islands

BROWSE ABOUT **GENOME UPLOAD** DOWNLOAD RESOURCES CONTACT US FAQ ACKNOWLEDGEMENTS HTTP API

DOWNLOAD RESOURCES CONTACT US FAQ ACKNOWLEDGEMENTS HTTP API LOGIN

GENOME UPLOAD

Users can log into the **user management interface** (see **LOGIN** menu) using Github or Google credentials facilitating access to results.

To analyze larger batches of genomes, an **HTTP API** is available to submit genomes and retrieve results programmatically using curl, python or perl. More information is available [here](#).

Genomic island predictions can be calculated for your genome using IslandPick, IslandPath-DIMOB, and SIGI-HMM. [More Information](#). [See our FAQs](#).

We only accept user genomes that are formatted in EMBL or GENBANK format. Examples of genome files you should submit: [GENBANK](#), [EMBL](#). If your data was generated by Prokka, please ensure you use Prokka's --complaint flag to generate standard compliant output. IslandViewer will not be able to process it otherwise.

Genbank ▾ Choose File No file chosen

Genome name (optional):

Email address to be notified upon completion (optional):

Upload (this may take several minutes)

Optional

Note for incomplete genomes

Incomplete genomes are now accepted as input due to popular demand. However, whole genome analysis is still preferred to reduce false predictions and missing GIs. Please use at your own risk. We suggest to minimize the number of contigs before performing GI analysis and carefully evaluate all results from such custom jobs. Contigs will be ordered against a user-selected reference genome using the Mauve contig orderer ([Rissman et al., 2009](#)). Please note that annotation of the contigs is still required.

IslandViewer 4: Job Upload

1. Shows the pipeline steps (Prepare → Distance → SIGI-HMM → IslandPick → IslandPath-DIMOB → Annotation → Validation)
2. You can refresh the page or add an email to get notified when the run is complete (it make take some minutes)

Meaning: This page only tells you the analysis progress. Final genomic island predictions appear once all steps are complete

IslandViewer 4 | An integrated interface for computational identification and visualization of genomic islands

BROWSE ABOUT GENOME UPLOAD DOWNLOAD RESOURCES CONTACT US FAQ ACKNOWLEDGEMENTS HTTP API LOGIN

The analysis is not yet complete, please check back to this page in a while, or you will be notified by email when the analysis is complete if you elected to enter an email address.

The graph to the right will show you the status of the Islandviewer pipeline.

Legend:

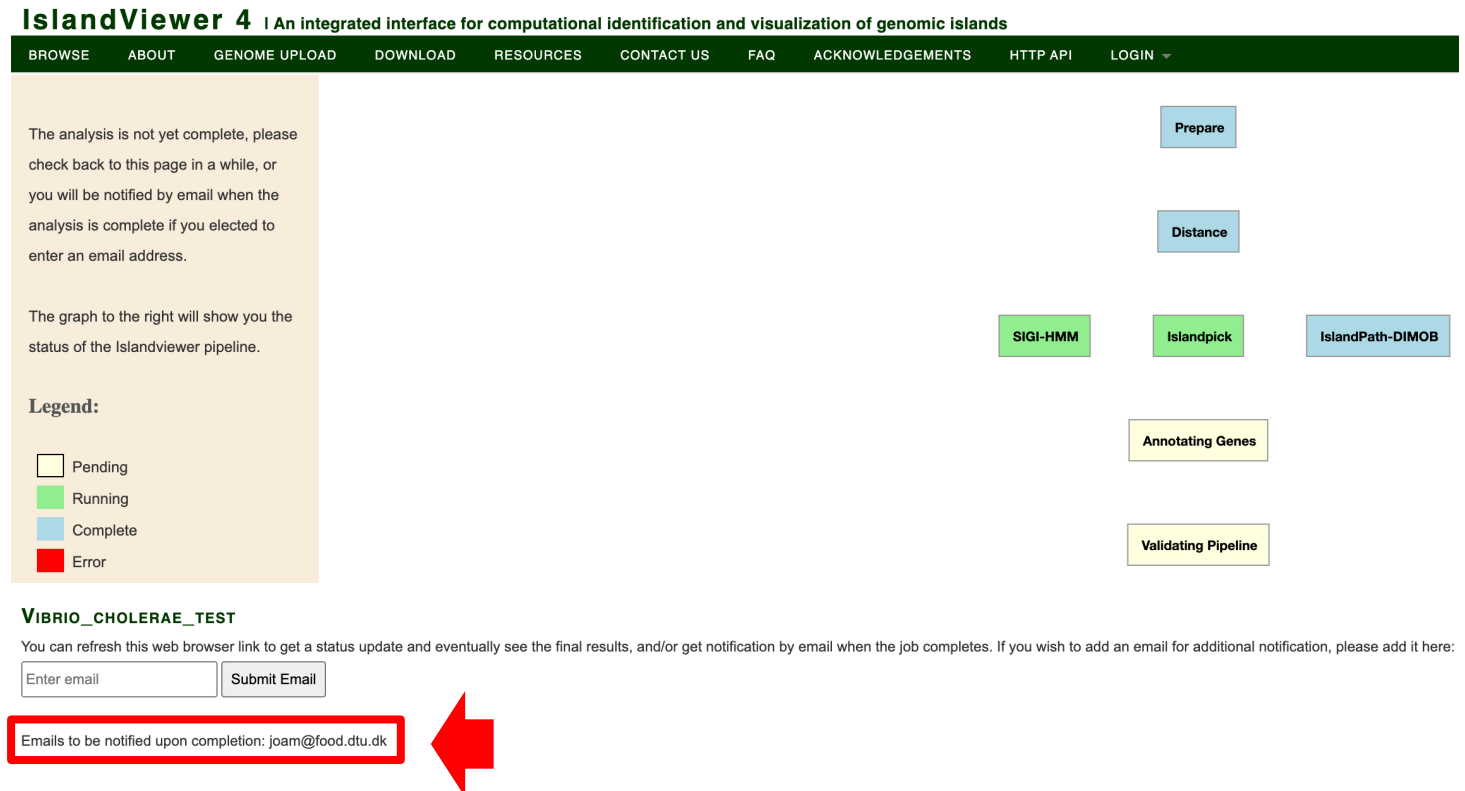
- ☐ Pending
- ☒ Running
- ☐ Complete
- ☐ Error

VIBRIO_CHOLERAE_TEST

You can refresh this web browser link to get a status update and eventually see the final results, and/or get notification by email when the job completes. If you wish to add an email for additional notification, please add it here:

Enter email Submit Email

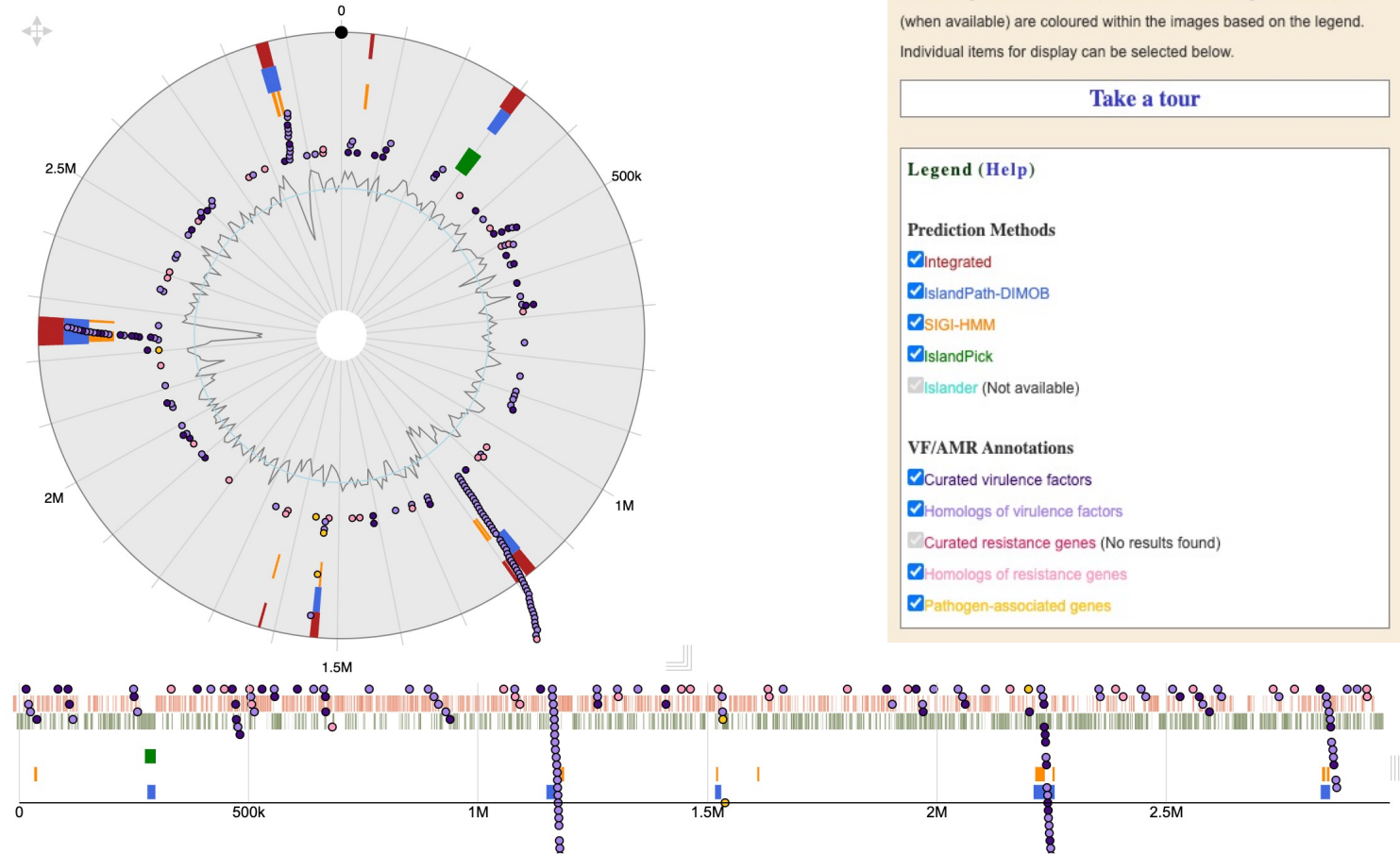
Emails to be notified upon completion: joam@food.dtu.dk



IslandViewer 4: Analysis of Results

1. **Circular map** (top): Chromosome with predicted genomic islands (colored blocks).
2. **Linear map** (bottom): Same information, but easier to zoom
3. **Colors** = methods used:
 - **Red** = integrated predictions (agreement between methods)
 - **Blue** = IslandPath-DIMOB
 - **Orange** = SIGI-HMM
 - **Green** = IslandPick
 - **Purple/red dots**: annotated virulence or resistance genes (if present)

VIBRIO CHOLERAE 2740-80 CHROMOSOME 1, COMPLETE SEQUENCE.

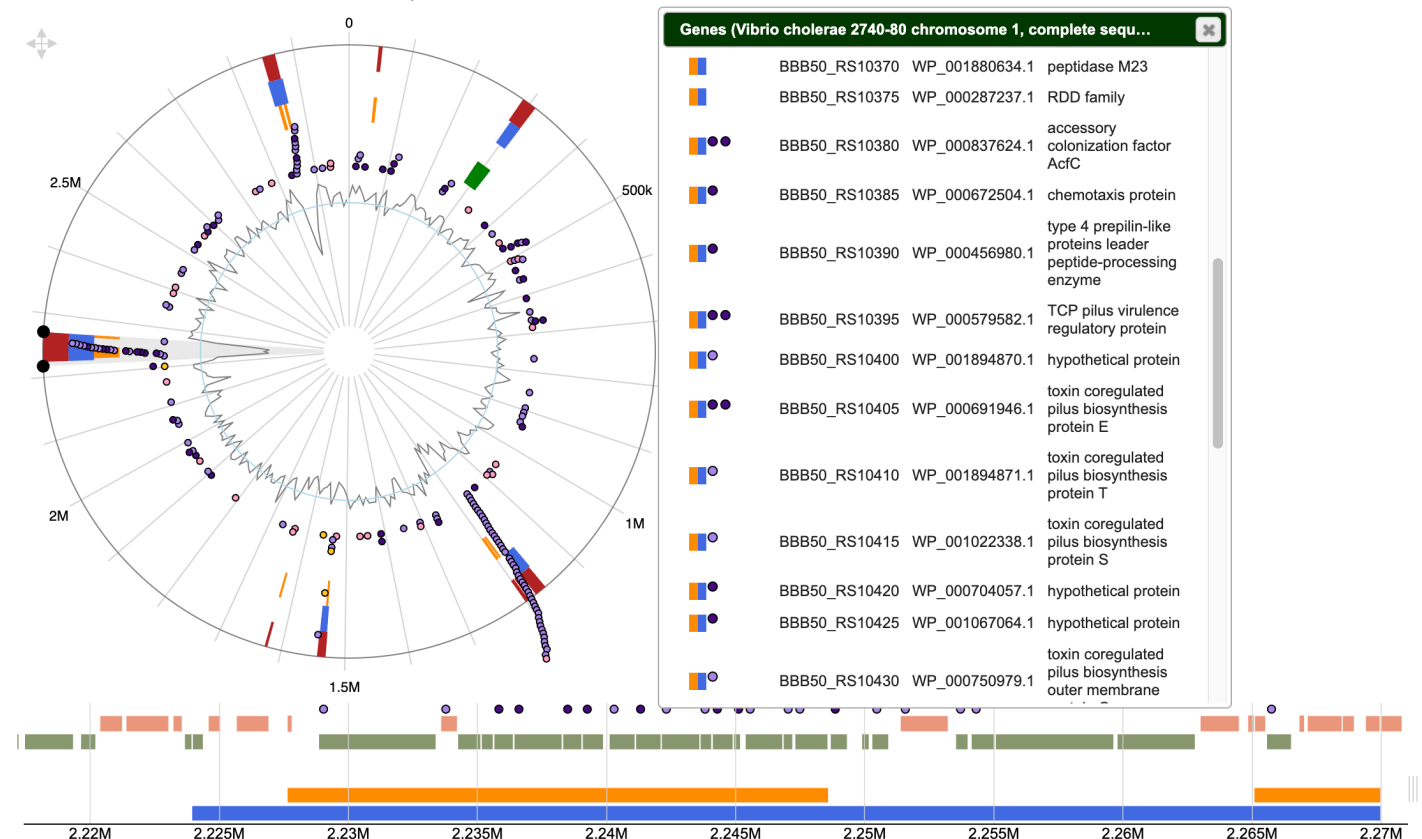


IslandViewer 4: Analysis of Results

1. By clicking in the **region** (≈ 2.23 – 2.25 Mb), you can see the genes inside that island
2. The **list shows**:
 - Method flags used to identify the GIs
 - Gene ID/locus tag
 - Protein name/function
3. No antimicrobial resistance genes were identified here

Meaning: This island likely contributes to pathogenicity rather than AMR

VIBRIO CHOLERAEE 2740-80 CHROMOSOME 1, COMPLETE SEQUENCE.



IslandViewer 4: Analysis of Results

1. The table lists all **putative genomic islands (GIs)** detected
2. Each **row** = one predicted GI with:
 - Start/End coordinates and size
 - Which **method(s)** flagged it (e.g. IslandPick, SIGI-HMM, etc.)
 - Click "**View**" to see region on the plot
 - **External annotations:** genes of interest inside the GI (e.g. virulence genes, pathogen-associated genes)
 - **Download links** for sequence, genes, and proteins

Start	End	Size	GI Prediction Method	View	External Annotations	Download (all data)
46,238	52,477	6,239	Predicted by at least one method			Sequence, Genes, Proteins
287,443	311,170	23,727	Predicted by at least one method			Sequence, Genes, Proteins
292,783	310,359	17,576	Predicted by at least one method			Sequence, Genes, Proteins
1,162,634	1,181,481	18,847	Predicted by at least one method		Virulence gene	Sequence, Genes, Proteins
1,190,190	1,194,472	4,282	Predicted by at least one method		Virulence gene	Sequence, Genes, Proteins
1,195,470	1,201,132	5,662	Predicted by at least one method		Virulence gene	Sequence, Genes, Proteins
1,530,047	1,543,566	13,519	Predicted by at least one method		Virulence gene, Pathogen Associated gene	Sequence, Genes, Proteins
1,532,443	1,536,681	4,238	Predicted by at least one method		Virulence gene, Pathogen Associated gene	Sequence, Genes, Proteins
1,622,122	1,626,499	4,377	Predicted by at least one method			Sequence, Genes, Proteins
2,224,200	2,270,213	46,013	Predicted by at least one method		Virulence gene	Sequence, Genes, Proteins
2,227,893	2,248,826	20,933	Predicted by at least one method		Virulence gene	Sequence, Genes, Proteins
2,265,338	2,270,213	4,875	Predicted by at least one method		Virulence gene	Sequence, Genes, Proteins
2,850,054	2,870,627	20,573	Predicted by at least one method		Virulence gene	Sequence, Genes, Proteins
2,853,051	2,859,864	6,813	Predicted by at least one method		Virulence gene	Sequence, Genes, Proteins
2,863,444	2,869,227	5,783	Predicted by at least one method		Virulence gene	Sequence, Genes, Proteins
287,443	311,170	23,727	IslandPick			Sequence, Genes, Proteins
46,238	52,477	6,239	SIGI-HMM			Sequence, Genes, Proteins

IslandViewer 4: Analysis of Results

1. IslandViewer predicts putative genomic islands (GIs) but **does not assign family names**
2. In *Vibrio cholerae*, naming major families (e.g., SXT/R391 ICEs, VPI-1/2) requires **alignment to known, curated GI** reference sequences (plus identification of hallmark genes)

 IslandCompare (within IslandViewer) currently provides curated GI references mainly for *Salmonella*, not *Vibrio*

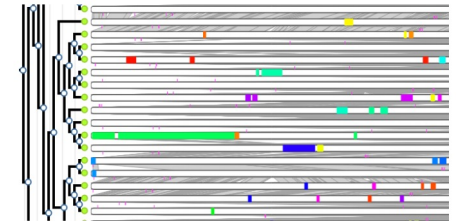
IslandCompare ANALYSE JOB HISTORY ABOUT FAQ DOWNLOAD PUBLICATIONS CONTACT

2025-03-18 - This application is currently unavailable as it undergoes maintenance/assessment. Please contact us at islandpick-mail@sfu.ca if you have any urgent needs. We apologize for any inconvenience.

IslandCompare (v1.1)

Genomic island prediction software developed to facilitate the analysis of microbial population datasets. IslandCompare is designed to process sets of microbial genomes and present genomic island content with an interactive visual designed to enable exploration of cross-genome genomic island content.

 Run Analysis



Getting Started

If you would like to test IslandCompare, please feel free to [view the pre-computed results of *Listeria monocytogenes* genomes](#) or download the [sample dataset](#). Alternatively, you can also [view a pre-computed set of *Pseudomonas aeruginosa* genomes](#). This second dataset also demonstrates how the alignment visuals allow you to see quickly see large structural changes in the genome, such as large genomic inversions.

Please note that Safari has known issues that will cause problems using this site. Please consider switching to Chrome or Firefox web browsers.

 View Example

 Tutorial

News and Updates

Thu Sep 22 2022

Curated Genomic Islands

IslandCompare is now adding support to detect curated, known *Salmonella enterica* genomic islands. In the visualization you will see them listed as the "Curated Genomic Islands" predictor. Additionally, if you hover your mouse over the island, it will show the genomic island name detected. If you are analyzing *Salmonella enterica* genomes in IslandCompare and want to see curated GI predictions for your genomes, make sure that the "Organism" field in your submitted file is filled out and includes the text "*Salmonella enterica*".

A major update of the IslandCompare backend brings in a variety of new features to improve stability and handle larger volumes of traffic. These updates will be occurring around Sept 22-23, and you may experience intermittent connectivity issues.

If you have any questions or problems, please don't hesitate to contact us at islandpick-mail@sfu.ca and thank you for your understanding as we implement

[View more...](#)

VCGIDB: *Vibrio cholerae* Genomic islands

[VCGIDB](#)[VC-GI LIST](#)[GENOME LIST](#)[BLAST SEARCH](#)[HELP](#)

VCGIDB

Vibrio cholerae Genomic islands

VCGIDB is a predicted genomic island database for *Vibrio cholerae* using GI-Scanner, large scale comparative genomic method to predict genomic island. From huge number of qualified whole genomes, GI-Scanner calculated pairwise comparisons and detect putative complementary CDSs making continuous array. CDS arrays were updated database and only corrected GIs by comparing phylogenetic tree were shown on this site.

479 Genomic islands detected from 798 Genomes



2010EL-1786 Genomic Island list

Show entries

Search:

GI-designation	Genes in GI	contig_index	begin	end
VC-GI 3	14	1	1041238	1049943
VC-GI 5	11	1	2600611	2609628
VC-GI 6 (VPI-2)	51	1	1368824	1424032
VC-GI 8	4	2	987529	991654
VC-GI 9	8	1	1050771	1055902
VC-GI 10 (VPI-1)	30	1	350805	383880
VC-GI 14	8	1	1432499	1440764
VC-GI 15	76	1	84073	180468
VC-GI 16	5	2	304110	308638
VC-GI 17	12	1	2947311	2959816

Showing 1 to 10 of 25 entries

Previous

1

2

3

Next

VCGIDB: *Vibrio cholerae* Genomic islands

VCGIDB

VC-GI LIST

GENOME LIST

BLAST SEARCH

HELP

Vibrio cholerae Genomic islands

Show entries

Search:

GI-designation	Alias	Genomes	Genes in GI (median)	Antimicrobial Resistance Genes	Description/Functional Annotation
VC-GI 3	CTX	34	14		transposon
VC-GI 5	VSP-1	393	11		
VC-GI 6	VPI-2	64	51		carbohydrate,transposon
VC-GI 8	GI-5	77	4		transposon
VC-GI 10	VPI-1	413	30		HGT
VC-GI 14	GI-4	100	8		carbohydrate
VC-GI 16	GI-7	647	5		
VC-GI 19	GI-9	623	7		
VC-GI 22		707	5		
VC-GI 28		724	6		
VC-GI 1	GI-12	1	19		
VC-GI 2	GI-12	3	19		
VC-GI 4	GI-12	1	17		
VC-GI 11	GI-12	1	17		
VC-GI 12	GI-11	30	45		phage
VC-GI 15	SXT	44	65	floR, APH(6)-Id, APH(3'')-Ib, sul2, tet(C)	HGT,plasmid,transposon
VC-GI 17	VSP-2	165	22		transposon

Abbreviations:

- **VC-GI #** — "*Vibrio cholerae* Genomic Island"
- **VPI-1 / VPI-2** — *Vibrio* Pathogenicity Islands 1 & 2
- **VSP-1 / VSP-2** — *Vibrio* Seventh-Pandemic islands 1 & 2 (hallmarks of 7th-pandemic El Tor)
- **CTX** — the CTX ϕ prophage carrying ctxAB (cholera toxin)
- **SXT** — the SXT/R391 family ICE

In summary

List of learning points in this session:

1. Genome annotation tools help detect key antimicrobial resistance and virulence traits.
2. ICEfinder detects ICE-like elements, but family identification may require sequence alignment.
3. IslandViewer predicts where genomic islands are, but it does not name families.
4. In *Vibrio*, identification of known GIs or ICE (e.g., VPI-1, VSP-2, SXT) requires comparison with literature-curated references and alignment tools (e.g., BLAST, progressiveMauve).

Acknowledgements

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