



Nanopore Sequencing Technologies

Outbreak tracking of respiratory viruses

Guillaume Croville – ENVT
11/04/2025

Intended Learning Objectives

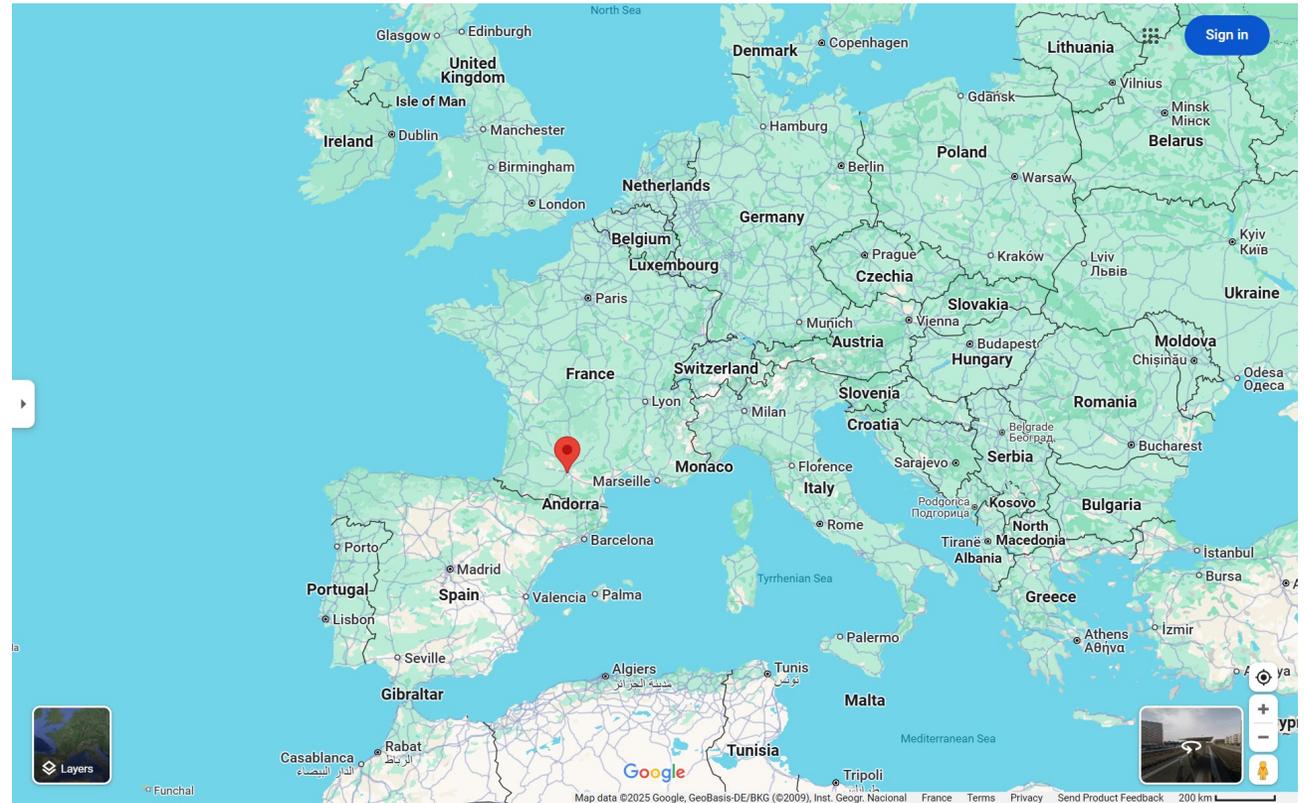
Specific objectives of this session:

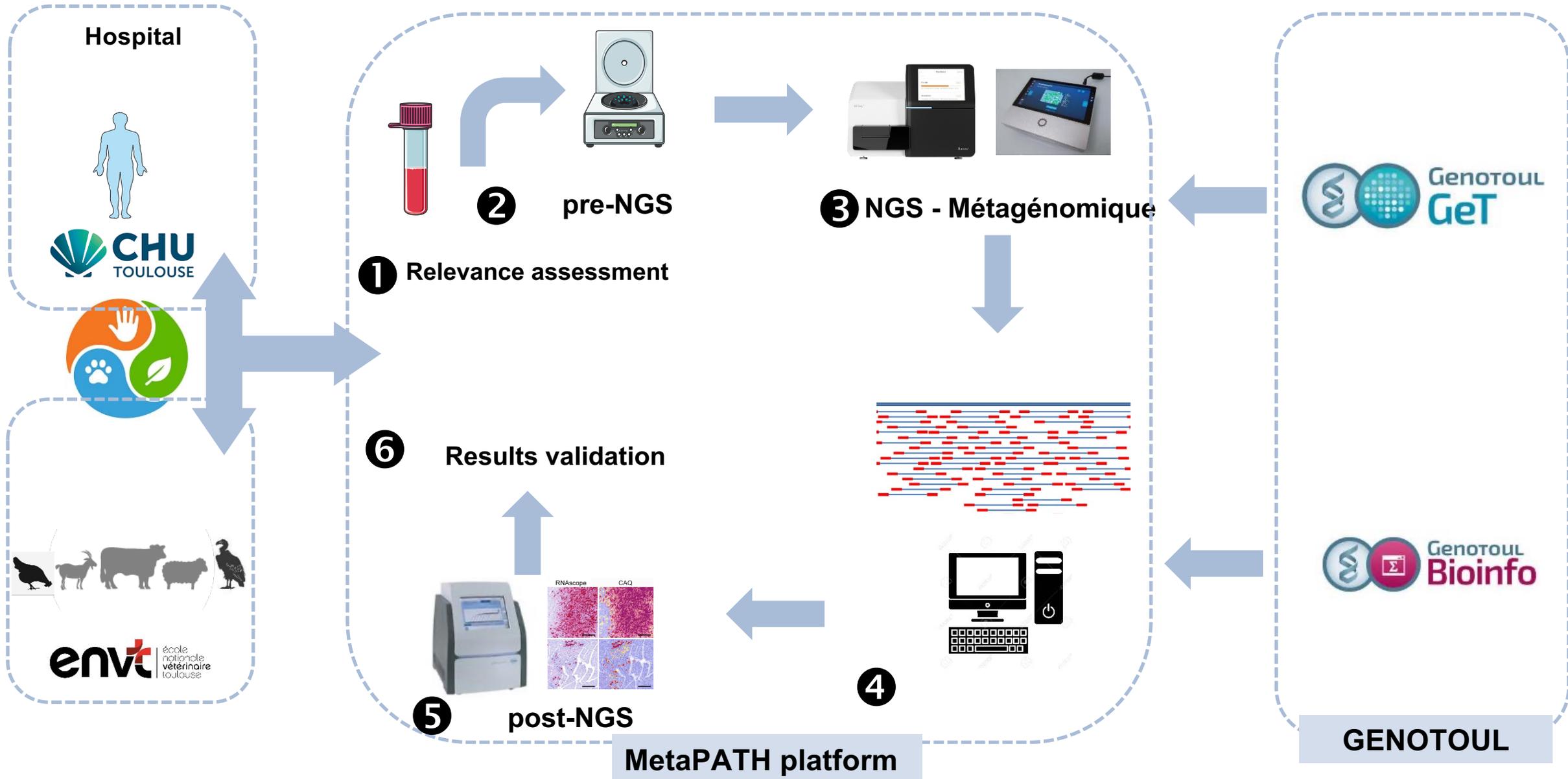
1. Learn about Nanopore sequencing applied to Influenza viruses
2. Learn about Nanopore sequencing for metagenomics

Outline

This session consists of the following elements

1. Presentation of an Influenza viruses sequencing study
2. Presentation of shotgun metagenomics studies



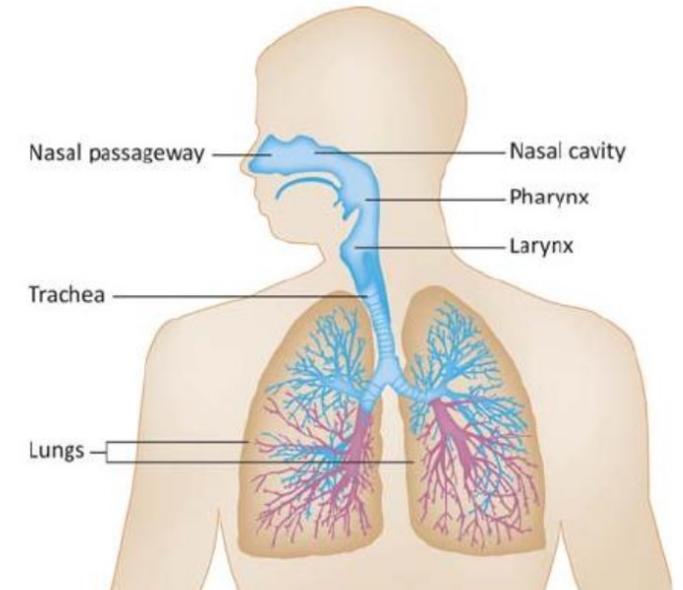
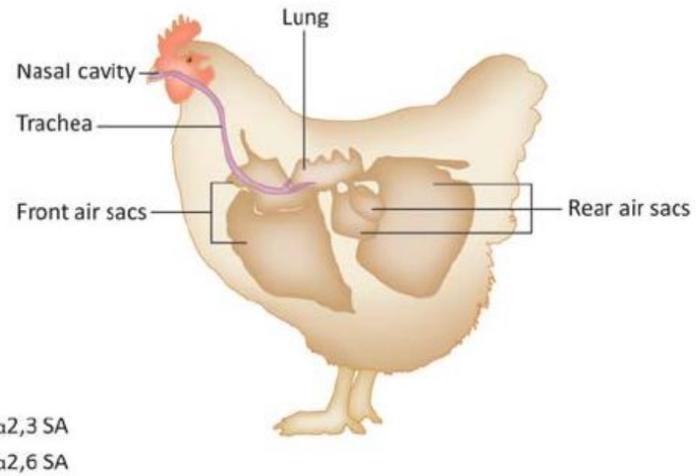
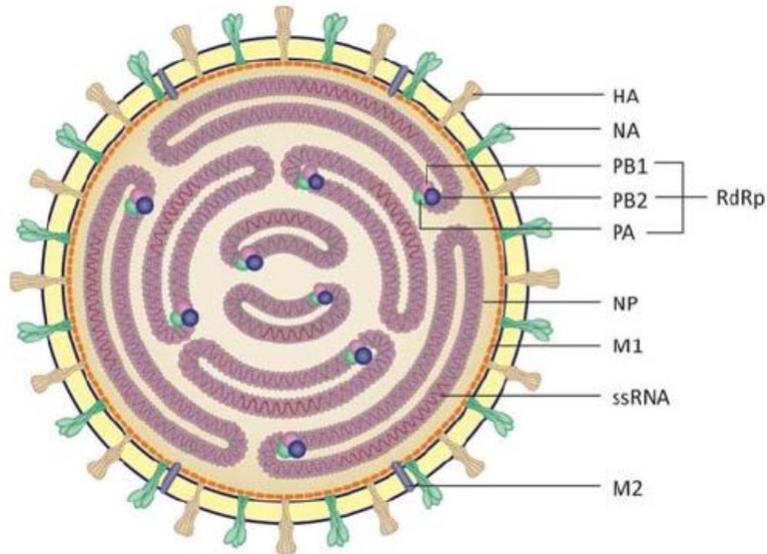


Outbreak tracking of respiratory virus in the field (example of the Avian flu)



Influenza virus genome is:

Avian influenza virus

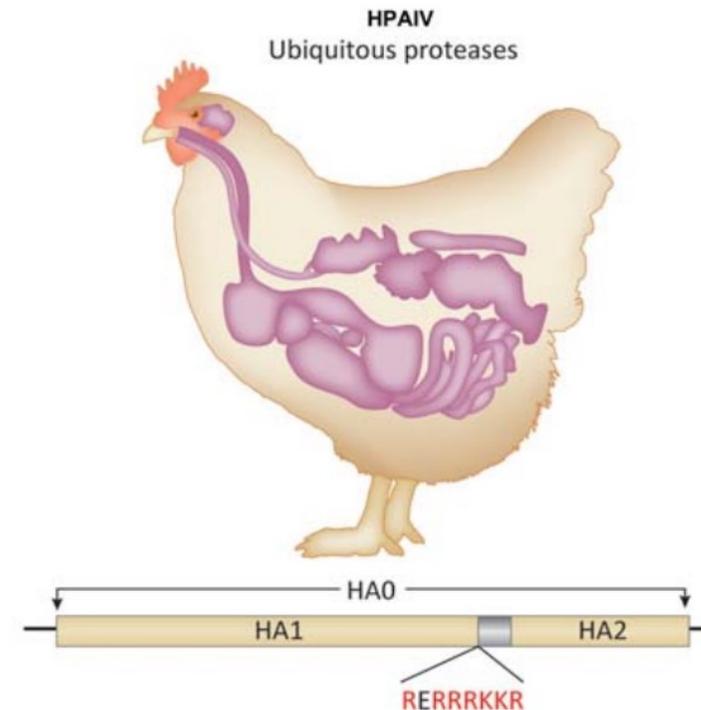
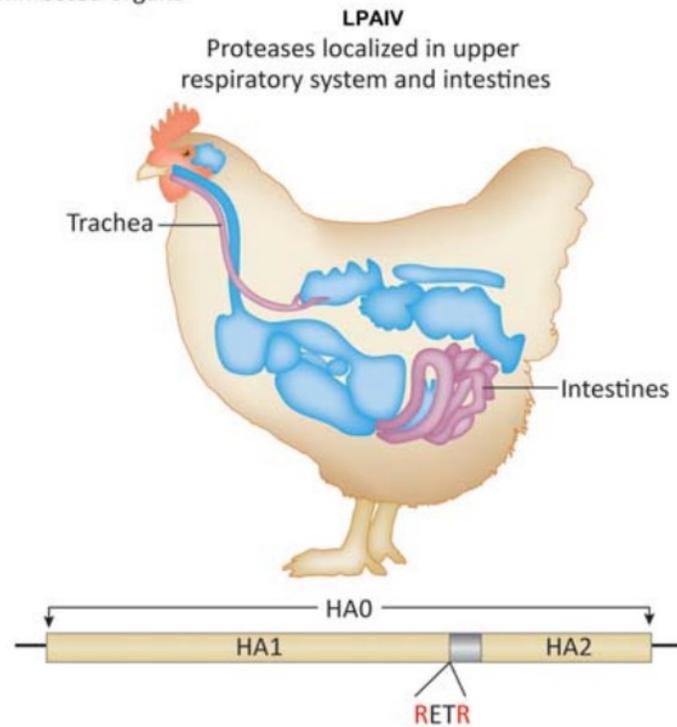


Susan Payne, in *Viruses*, 2017

Avian influenza virus

Tissue tropism of LPAIV and HPAIV

■ Infected organs
■ Uninfected organs



Susan Payne, in *Viruses*, 2017

Avian influenza in France, 2020-2021

 Veille Sanitaire Internationale
Plateforme ESA

Interactive application of visualization of the outbreaks and cases in Europe for the main diseases monitored by French Animal Health Epidemic Intelligence

Settings

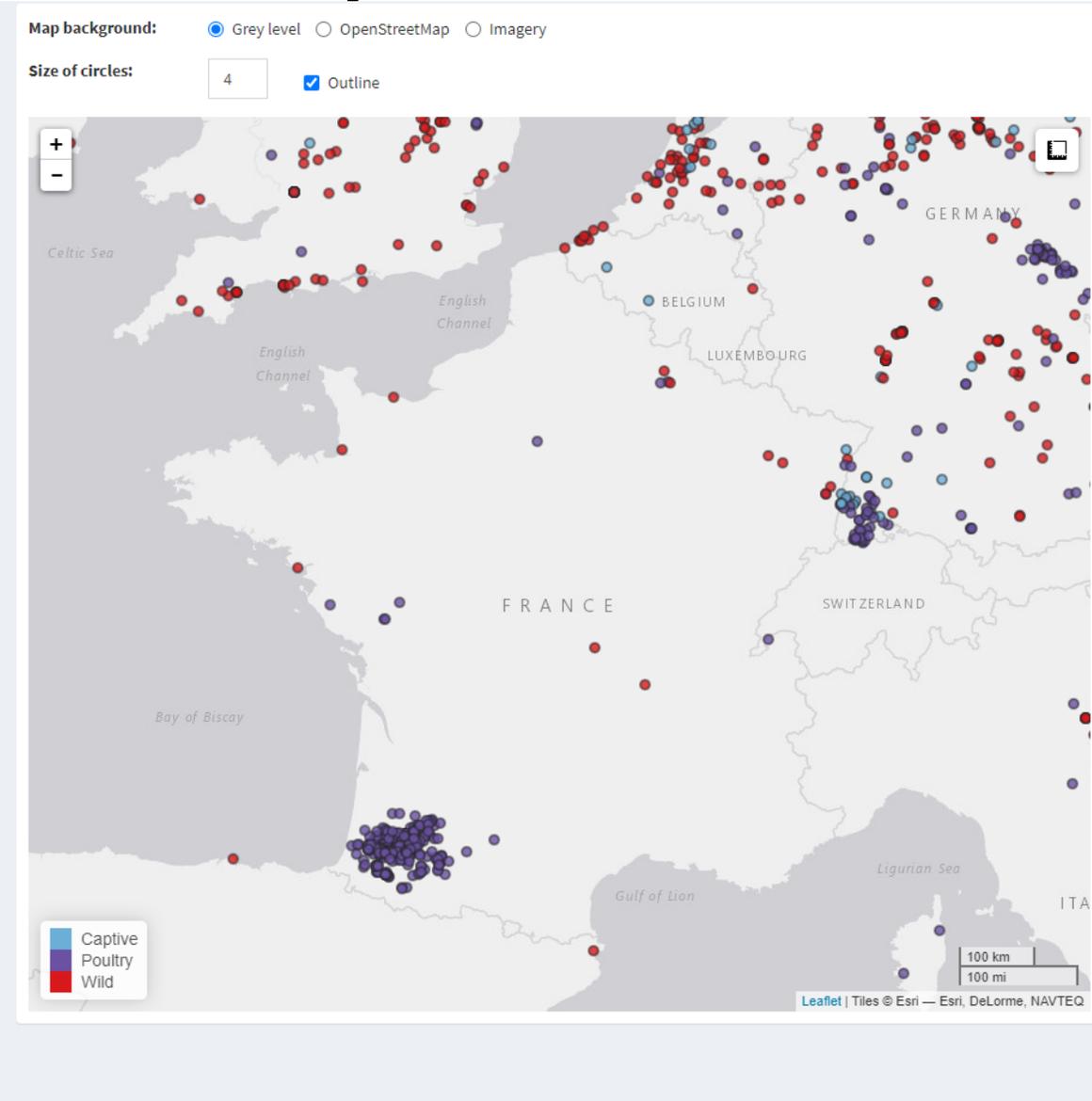
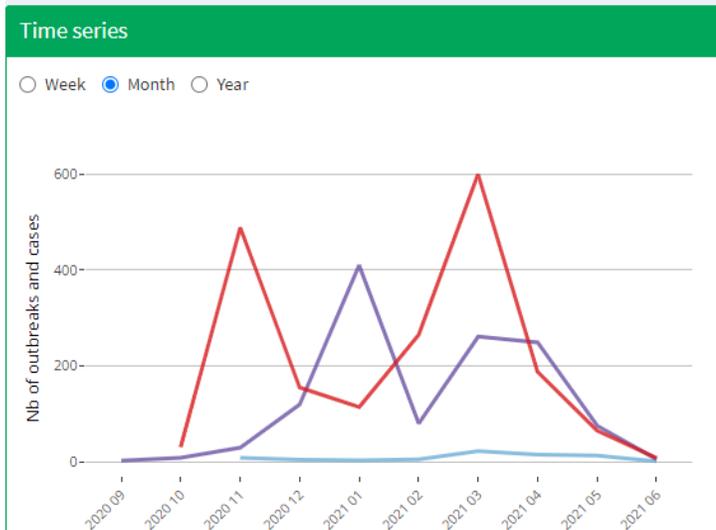
Select disease: Highly Pathogenic Avian Influenza

Period: 2018-10-26 — 2020-08-26 — 2021-06-05 Play video

Serotypes: H5N1 H5N2 H5N3 H5N4 H5N5
 H5N6 H5N8 H5Nx H7N7

Species: Wild Poultry Captive

Data source: ADIS / WAHIS
To obtain older data or to export figures, contact the [Epidemic Intelligence team](#)



Avian influenza in France, 2021-2022

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Settings

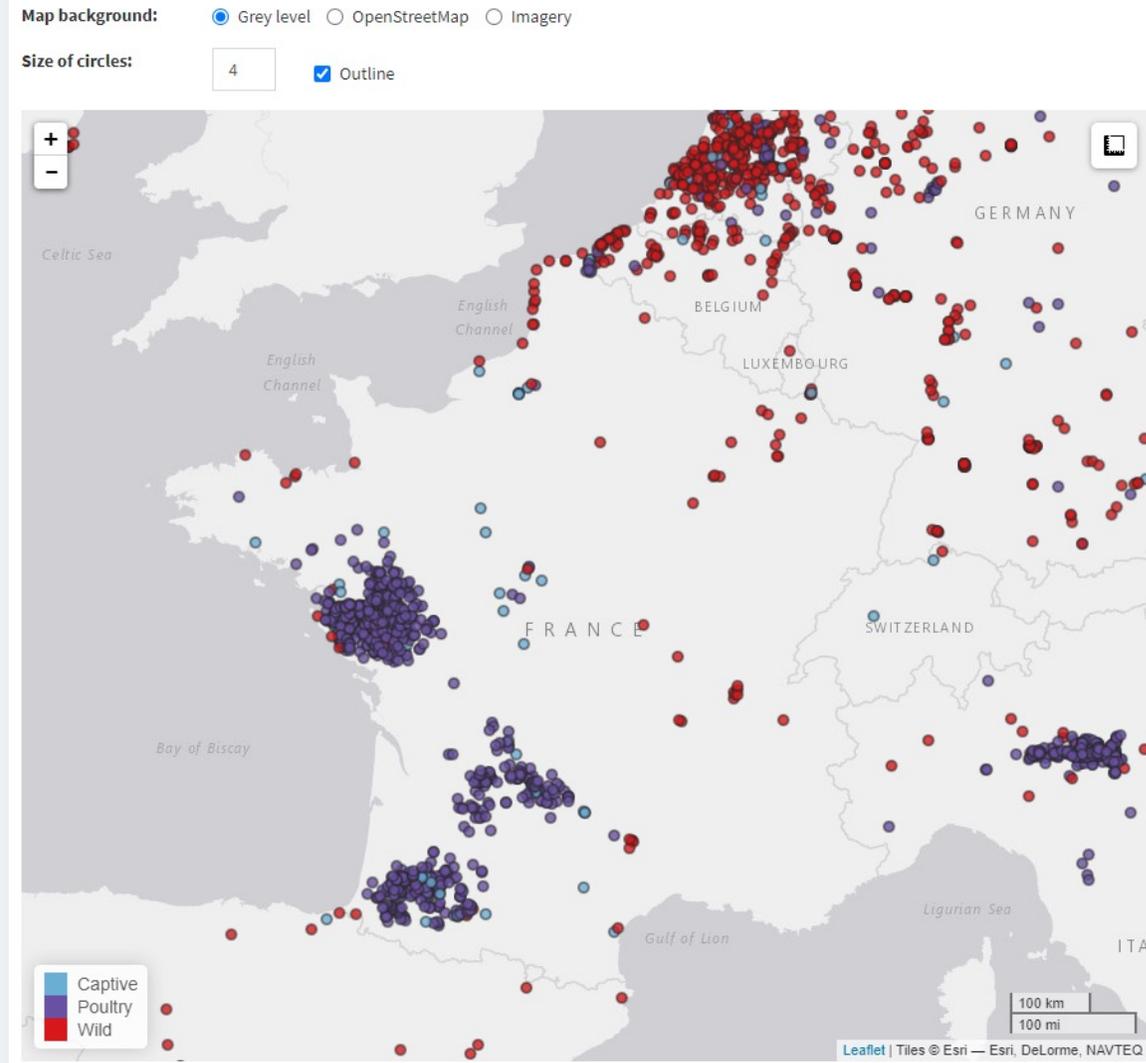
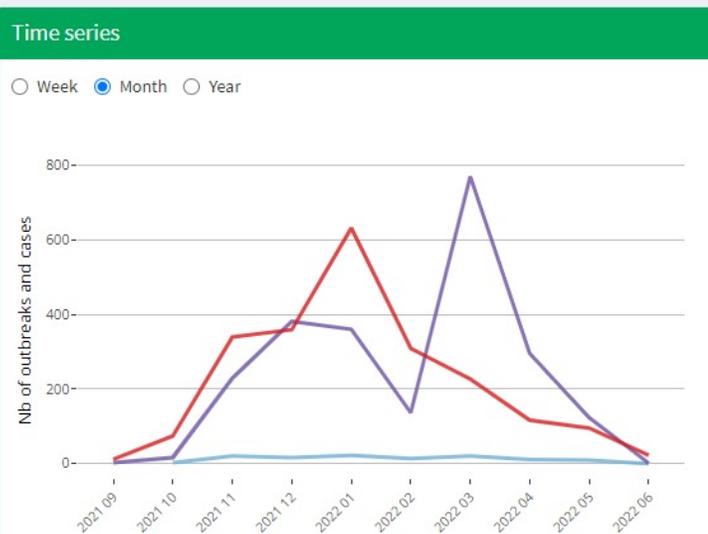
Select disease: Highly Pathogenic Avian Influenza

Period: 2018-10-28 — 2021-09-13 — 2022-06-05 Play video

Serotypes: H5N1 H5N2 H5N3 H5N4 H5N5
 H5N6 H5N8 H5Nx H7N7

Species: Wild Poultry Captive

Data source: ADIS / WAHIS
To obtain older data or to export figures, contact the Epidemic Intelligence team



Avian influenza in France, 2022-2023

 Veille Sanitaire Internationale
Plateforme ESA

Interactive application of visualization of the outbreaks and cases in Europe for the main diseases monitored by French Animal Health Epidemic Intelligence

Settings

Select disease:

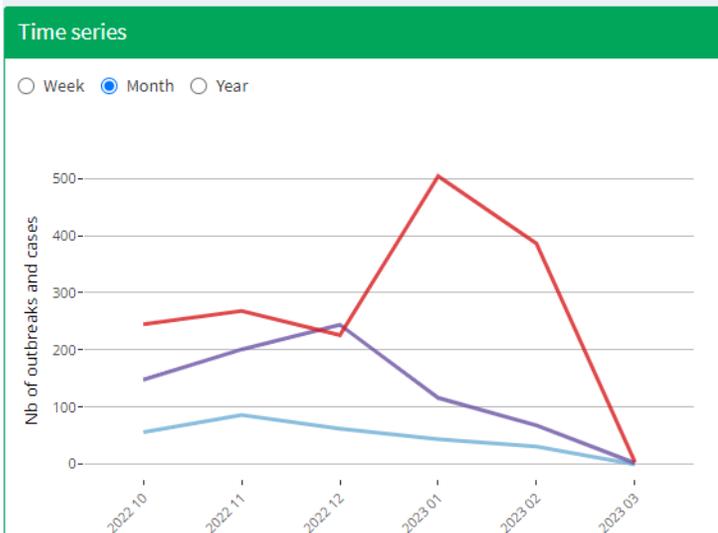
Period:

2016-10-26 2018-02-09 2019-05-27 2020-09-10 2021-12-26 2023-03-02

Serotypes: H5N1 H5N2 H5N3 H5N4 H5N5
 H5N6 H5N8 H5Nx H7N7

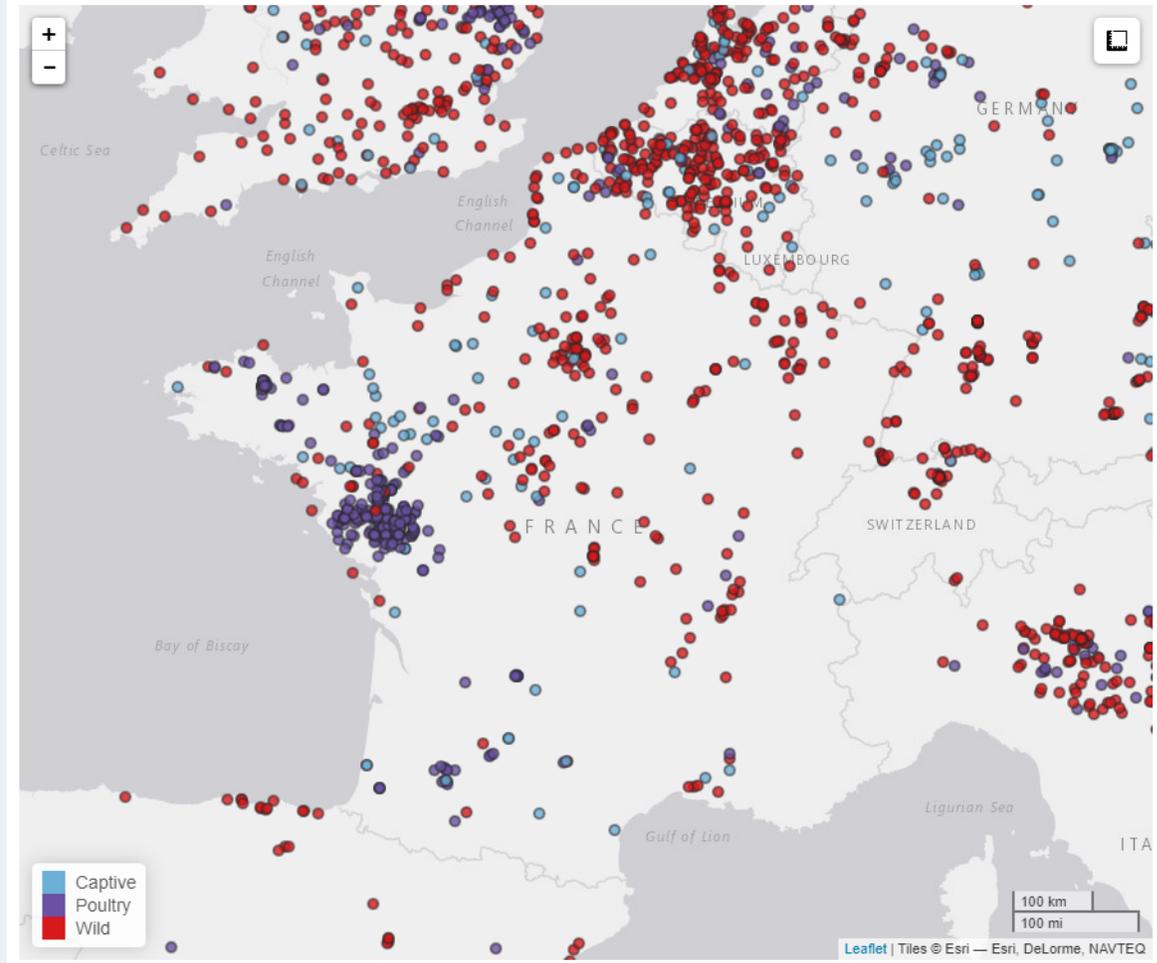
Species: Wild Poultry Captive

Data source: ADIS / WAHIS
To obtain older data or to export figures, contact the [Epidemic Intelligence team](#)



Map background: Grey level OpenStreetMap Imagery

Size of circles: Outline



Avian influenza in France, 2020-2023

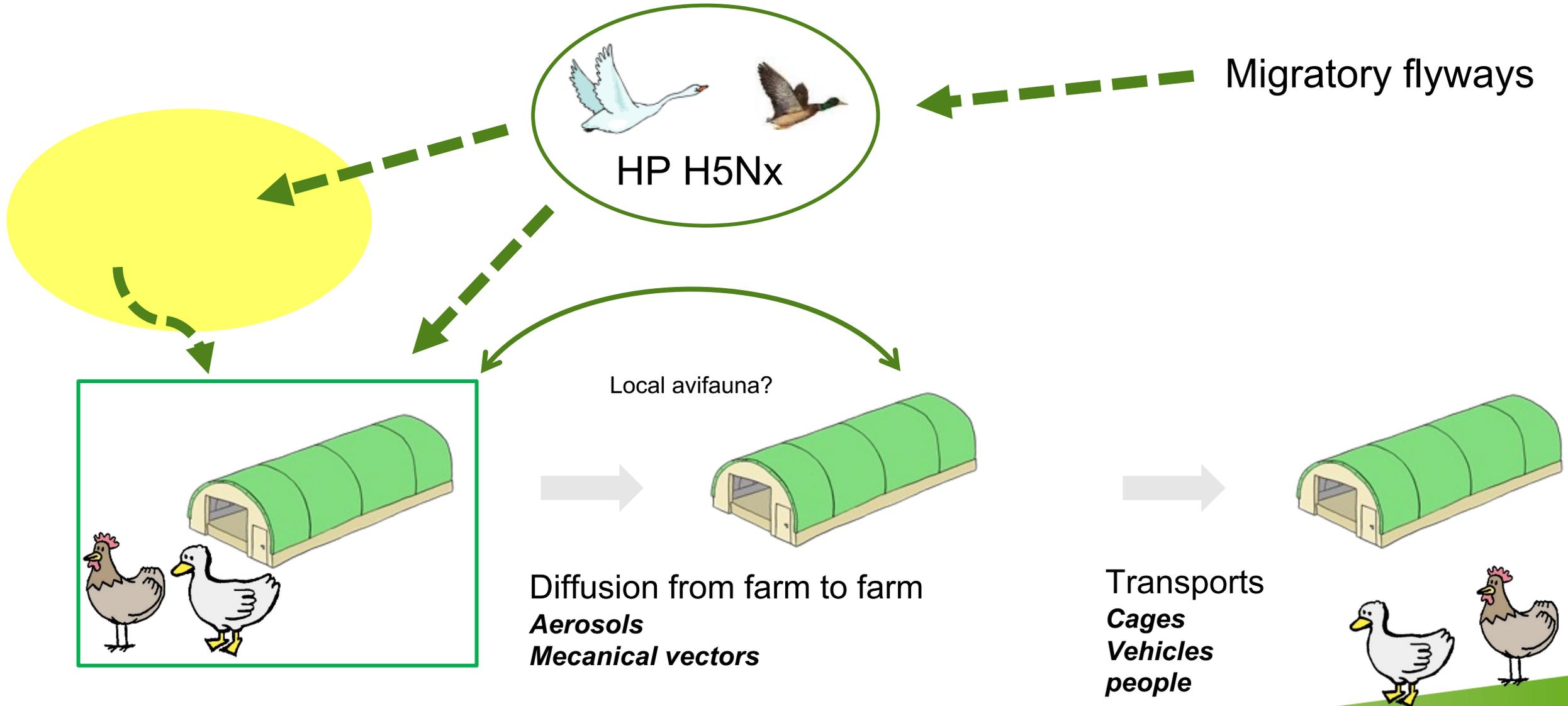
	2020-2021 (H5N8)	2021-2022 (H5N1)	2022-2023 (H5N1)
First outbreak	16 November 2020	26 November 2021	1st August 2022
Total outbreaks	514	1476	314

Questions to address during an influenza epizootic:

- Pathotyping (HP vs LP)
- Typing (HxNy)
- New introduction vs circulating strain
- ...

➔ Nanopore sequencing?

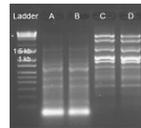
The spread of HPAIVs from wild birds to farms



Workflow

Day 1

- Sampling
- (On-field screening)
- RNA extraction
- Influenza real-time PCR
- Influenza mPCR

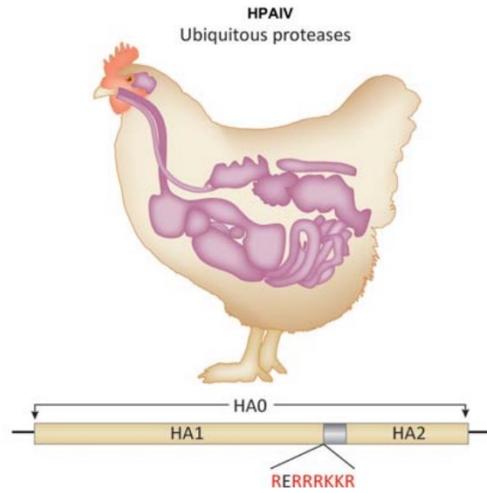


Day 2

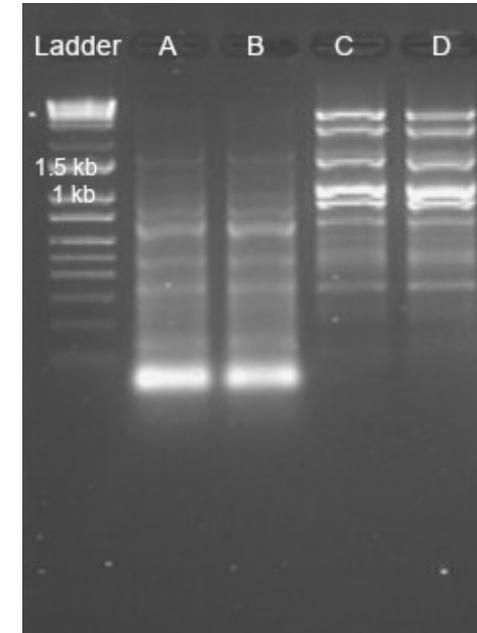
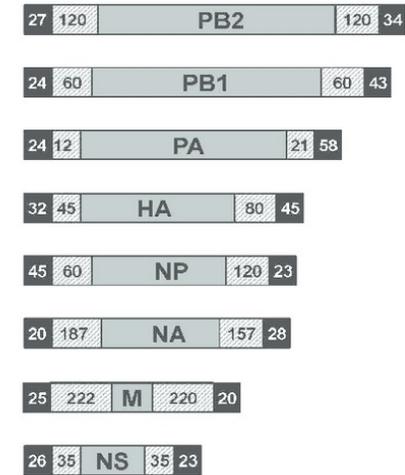
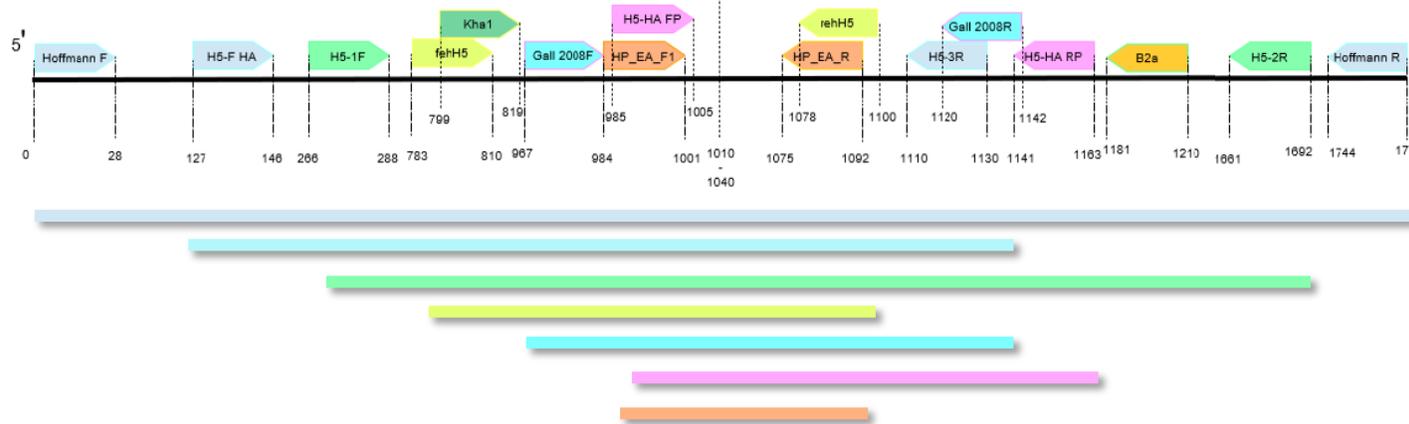
- Library prep
- Sequencing
- Bioinformatics



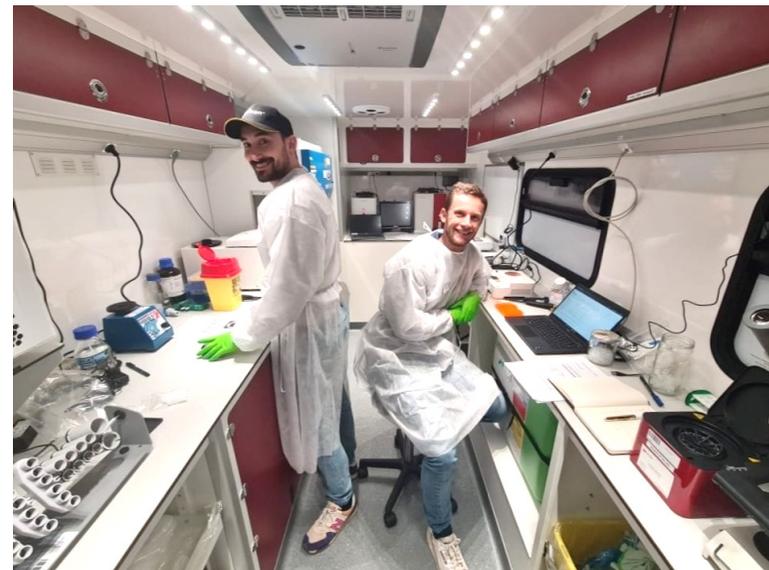
Influenza PCR



Cleavage site



A lab in a truck



Oxford Nanopore sequencing (3 samples)



Kit type: SQK-NBD112-24

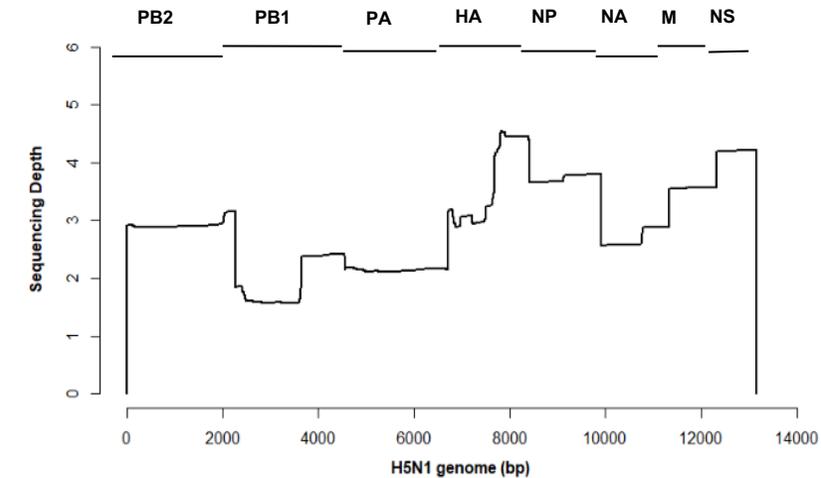
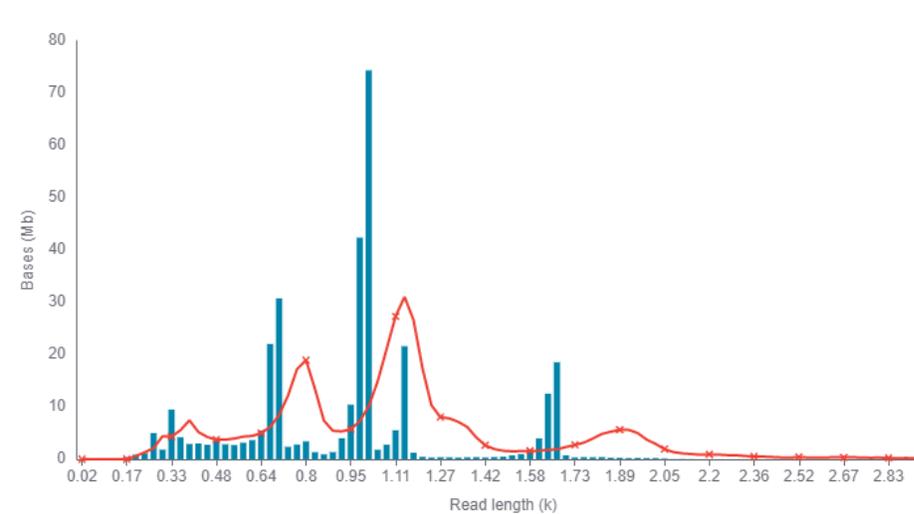
Run duration: 1 hour 45 minutes

Bases called: 331,72 Mb

Flow cell type: FLO-MIN106

Reads generated: 477.13 k

N50: 1,02 kb



Bioinformatics



minimap2 -ax map-ont reference.fa mysample.fastq > mysample.sam

samtools sort mysample.sam > mysample.sort.bam

samtools index mysample.sort.bam

samtools flagstat mysample.sort.bam > mysample_flagstat

samtools depth mysample.sort.bam > mysample_samdepth

samtools mpileup -A -Q 0 mysample.sort.bam | **iVar consensus -p prefix -q 10 -t 0 -m 1**

Minimap2: Li, H. 2018

Samtools: Li, 2011

iVar: Grubaugh *et al.* 2019

Avian influenza outbreaks tracking 2020-2022

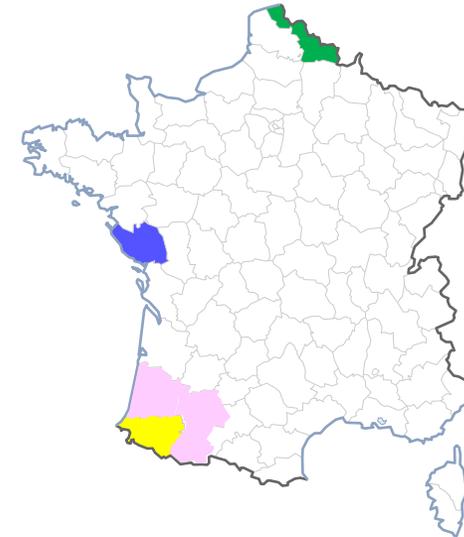
2020-2021 (H5N8)

- 11 samples : tracheal swabs, feathers, dust
- CT value : 14 to 28,5
- Flongle FC
- 28 to 474Mb per sample



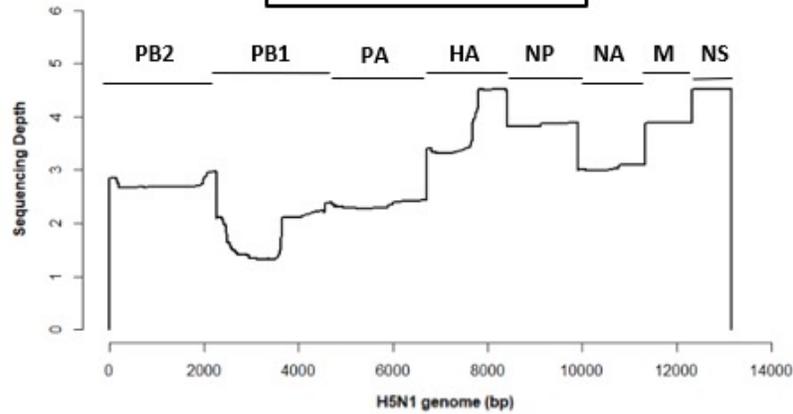
2021-2022 (H5N1)

- 19 samples : tracheal swabs, cloacal swabs feathers, dust
- CT value : 13 to 27
- Flongle FC
- 49 to 205Mb per sample

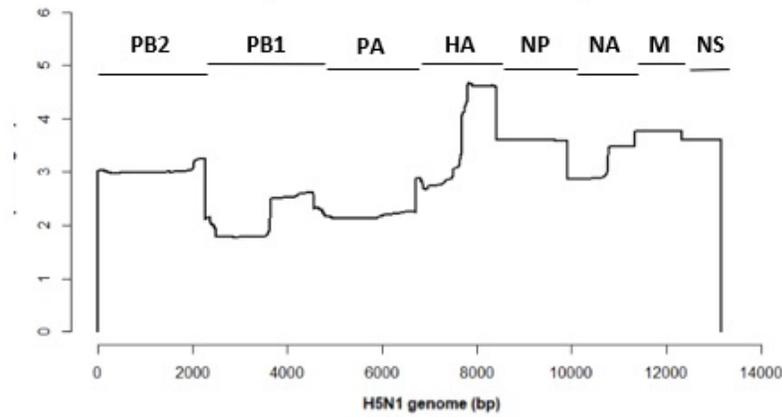


Outbreaks tracking 2020-2022

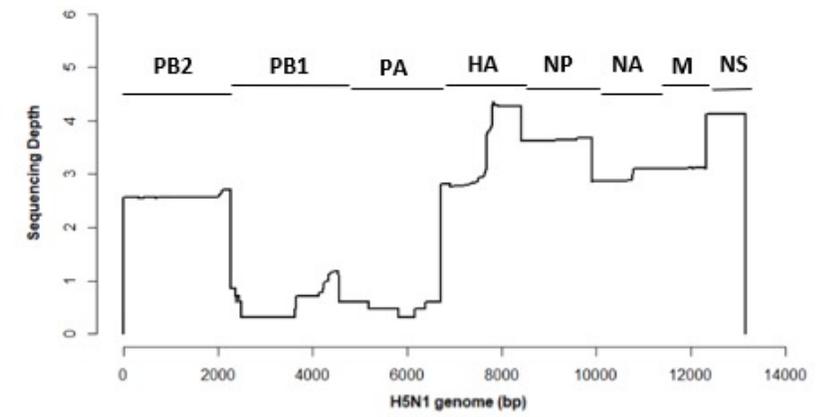
LAB21-328_Z1



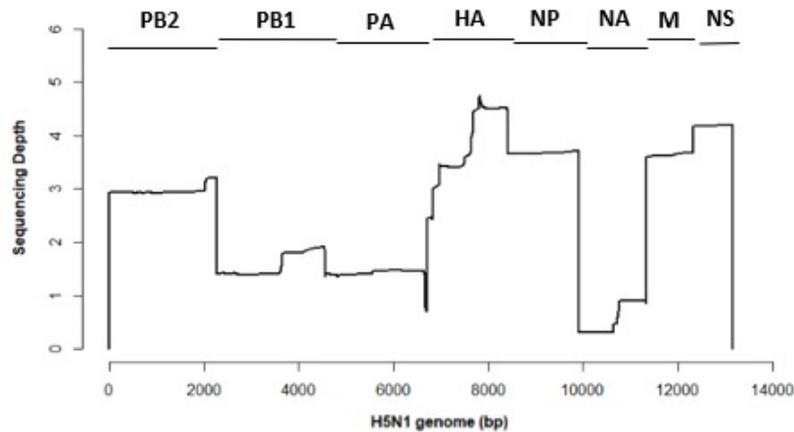
LAB21-328_Z7



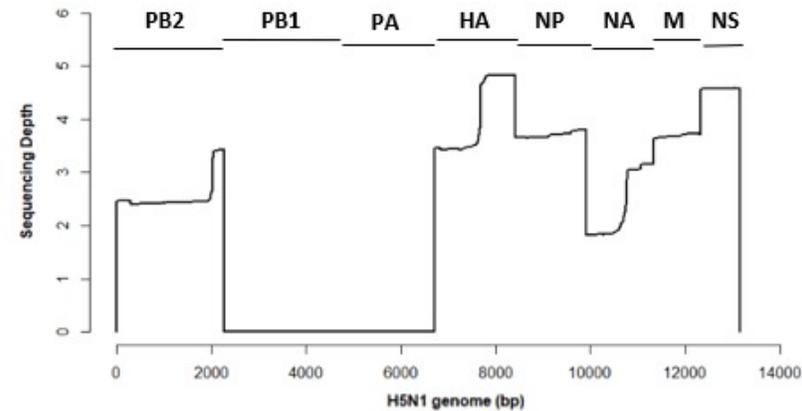
LAB21-329_Z7



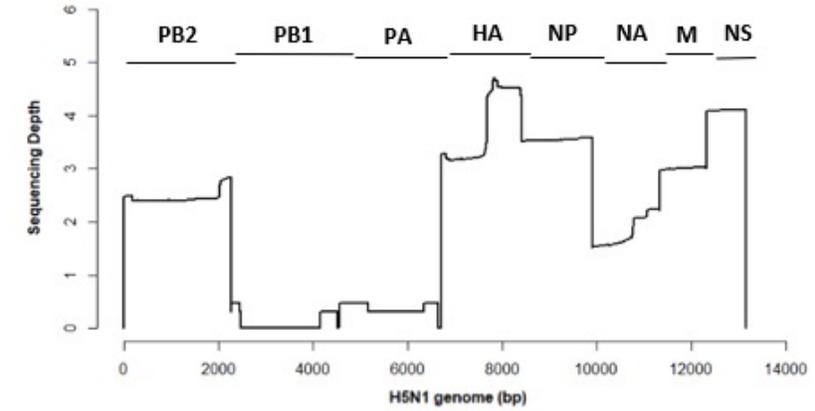
LAB21-343_P11



LAB21-347_P5



LAB21-347_P8



Outbreaks tracking 2020-2022

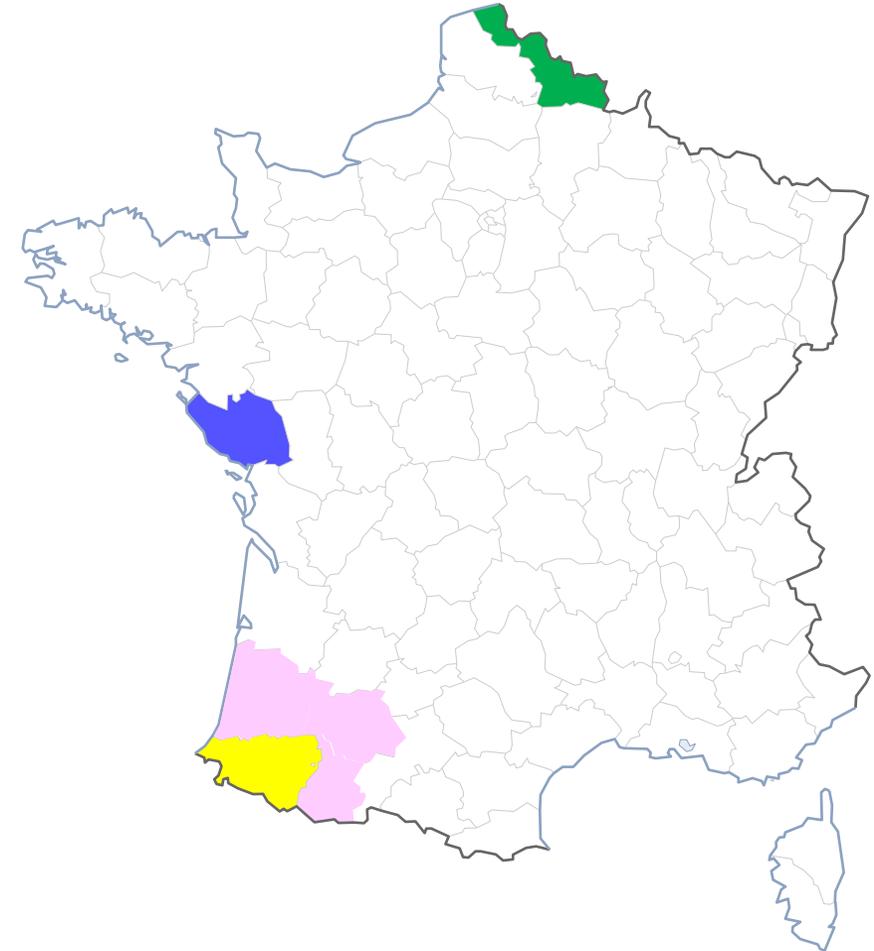
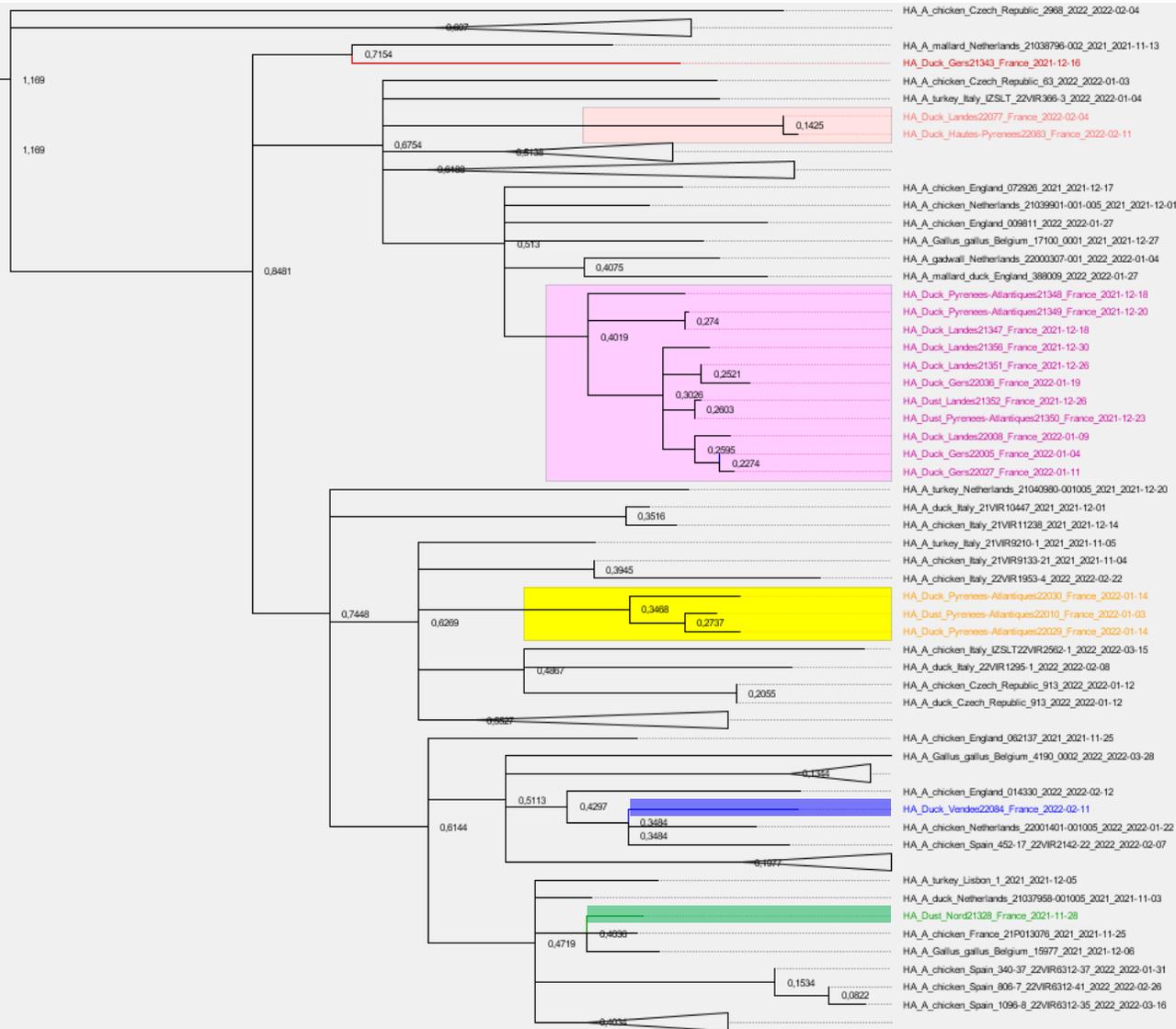
Pathotyping

HA/Duck/Landes20323/Franc	KLVLATGLRNNPIREKRRKRLGFGAIAO
HA/Duck/Landes20335/Franc
HA/Duck/Landes20349/Franc
HA/Duck/Vendee20338/Franc
HA/Duck/Deux-Sevres20339/R.....
HA/Duck/Landes20347/Franc
HA/Duck/Landes20352/Franc
HA/Duck/Landes20353/Franc
HA/Duck/Hautes-Pyrenees21
HA/Duck/Hautes-Pyrenees21
HA/Duck/Gers21084/France/

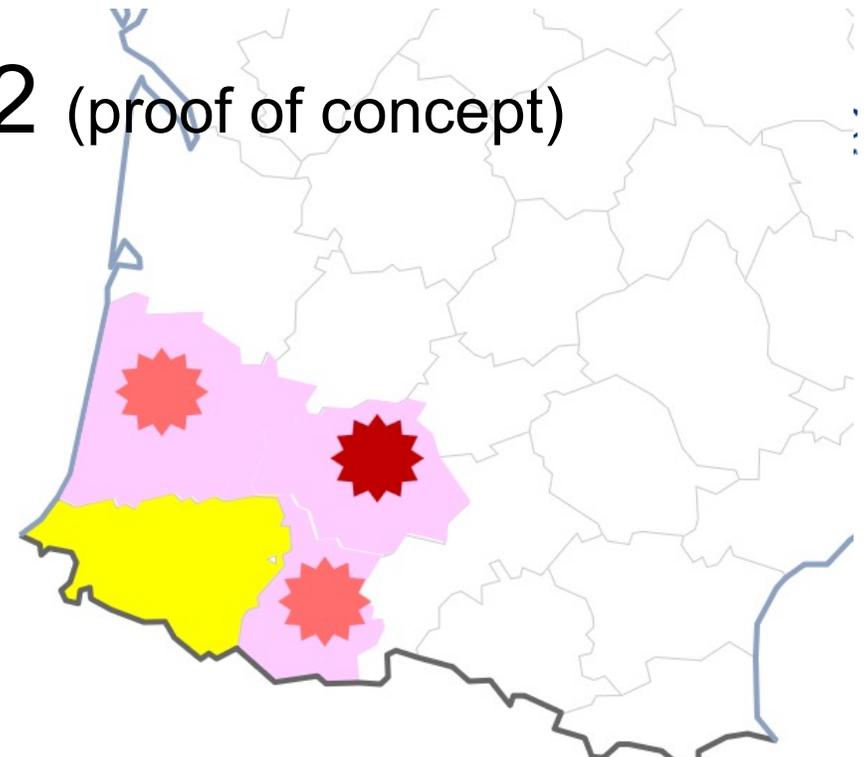
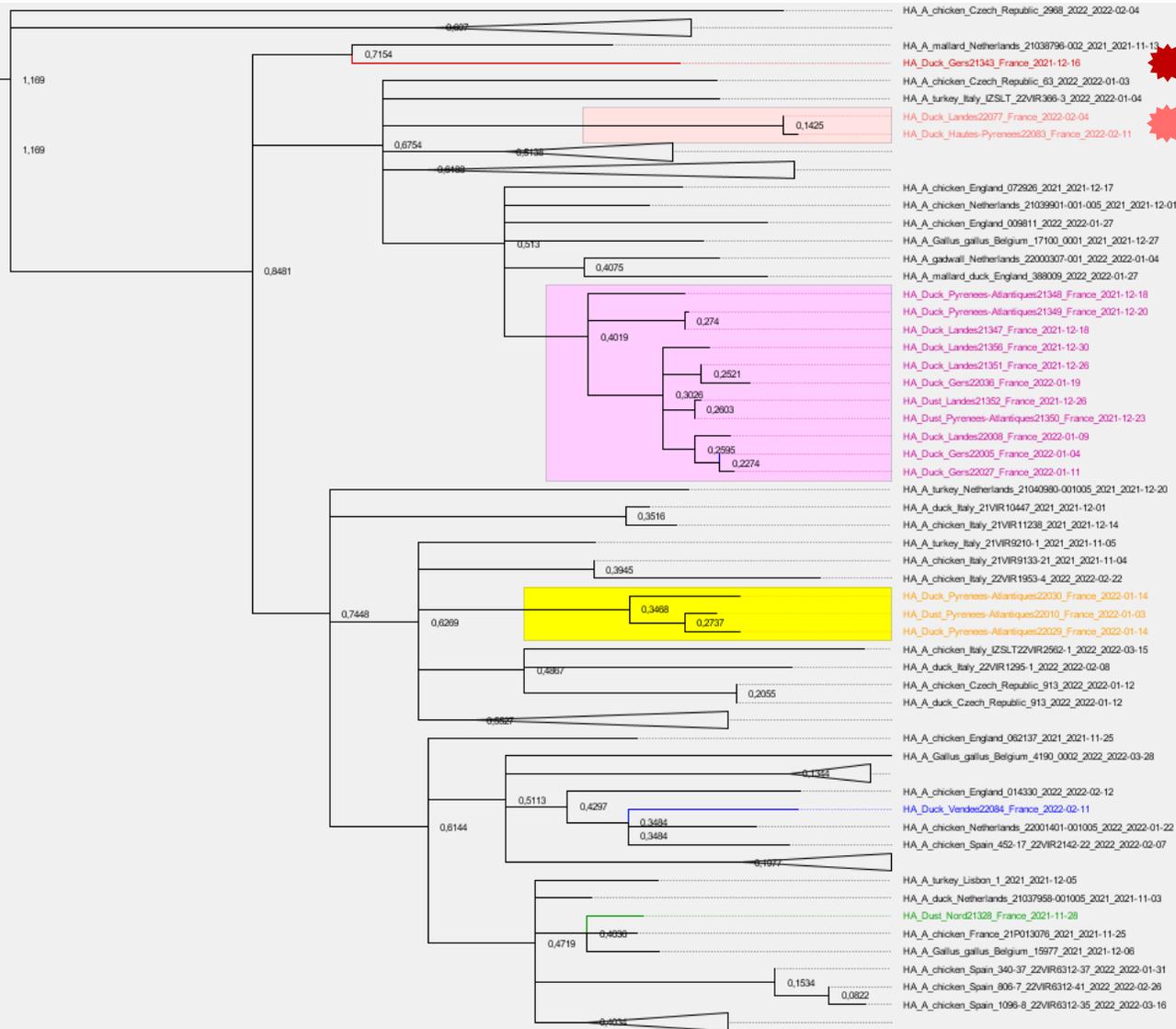
Identification of cluster specific mutations

HA/Duck/Landes20323/France/20201206	EWSYIVERANPANDLCYPGSLNDYEELKHLISRINHFEKILIIIPKSSWPNHETSLGVSAACPYQGAPSFRRNVVWLIKKNDAYPTIKISYNNNTNREDLLILWGIHHSNNAEEQ
HA/Duck/Landes20335/France/20201210
HA/Duck/Landes20349/France/20201221
HA/Duck/Vendee20338/France/20201213N.....R.....Q.....V.....D.A
HA/Duck/Deux-Sevres20339/France/20201213Q.....K.....N.....R.....Q.....V.....D
HA/Duck/Landes20347/France/20201215
HA/Duck/Landes20352/France/20201226
HA/Duck/Landes20353/France/20201227
HA/Duck/Hautes-Pyrenees21061/France/20210128R.....S
HA/Duck/Hautes-Pyrenees21064/France/20210130R.....N.....S
HA/Duck/Gers21084/France/20210220R.....S

Outbreaks tracking 2021-2022



Outbreaks tracking 2021-2022 (proof of concept)



→ Introductions (★) not linked with circulating strains

An amplicon-based nanopore sequencing workflow for rapid tracking of avian influenza outbreaks, France, 2020-2022.

Croville G, Walch M, Sécula A, Lèbre L, Silva S, Filaire F, Guérin JL.

Front Cell Infect Microbiol. 2024 Jan 22;14:1257586. doi: 10.3389/fcimb.2024.1257586. eCollection 2024.

PMID: 38318163 Free PMC article.

Conclusion

- Combination of mPCR with ONT:
 - Typing, pathotyping, phylogeny in <48h
 - Whole genome sequencing / good seq. depth
- MK1C 🥰
- (Expired) MinION Flowcell 🥰

Nanopore sequencing for metagenomic studies



Metagenomics is:

① The Slido app must be installed on every computer you're presenting from

Metagenomics

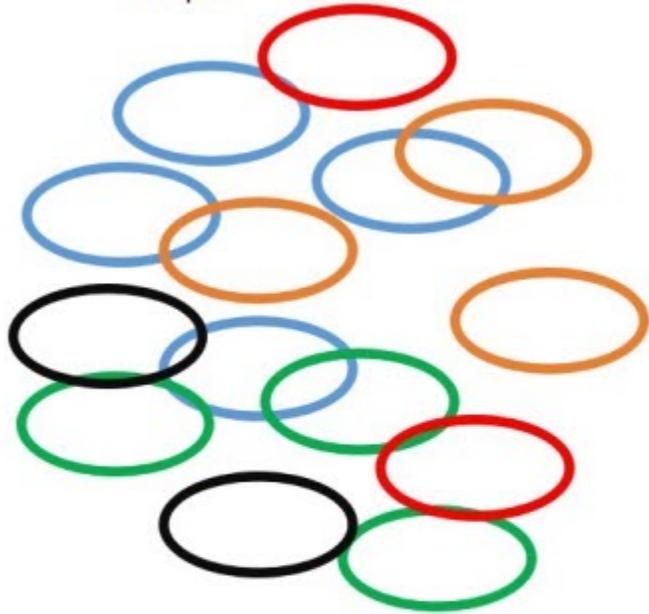
Metagenomics aims to study the **microbiome**.

Microbiome: the set of micro-organisms (bacteria, viruses, fungi, yeasts, plankton...) living in a specific environment (intestine, ocean, soil, air, etc.).

Metagenomic can be performed using 2 approaches:

- **Targeted metagenomics** or **metabarcoding**.
- **Shotgun metagenomics**.

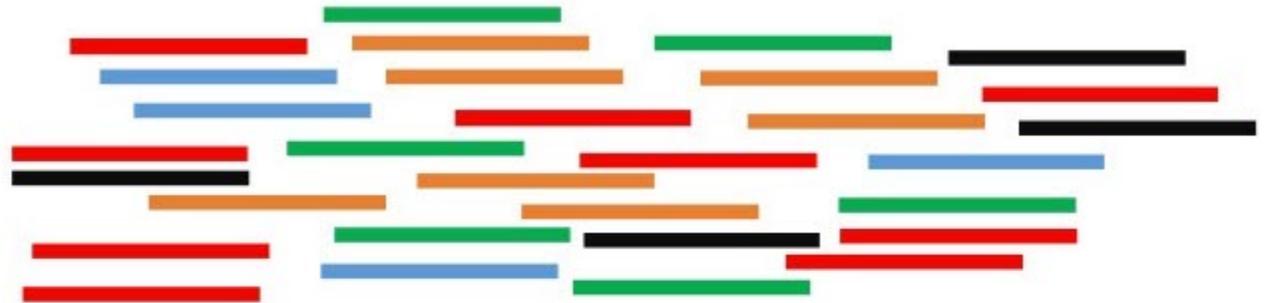
Genomes of the
microorganisms included in the
sample



DNA
Extraction



Fragmentation



Sequencing



Unknown genome
↑

Assembly and alignment with genomes referenced in
databases

Assembly and new
annotation

- 16 Max depth
- 11 Font size
- Chart size
- Color by Avg. score
- Collapse
-
-
-



- Eukaryota 0.2%
- Euryarchaeota 0.2%

Interactive metagenomic visualization in a Web browser

[Brian D Ondov](#), [Nicholas H Bergman](#) & [Adam M Phillippy](#)

ARN_22-1152R-30A	OTU.ID	Rank1	Rank2	Rank3	Rank4	Rank5	Rank6	Rank7
167037	11636	k_Viruses	p_Artvervir	c_Revtraviricetes	o_Ortervirif	f_Retrovirif	g_Gammaretrovirus	s_Reticuloendotheliosis virus
39362	1623289	k_Viruses	p_Uroviric	c_Caudoviricetes	o_	f_	g_Dhillonvirus	s_
18986	353768	k_Viruses	p_Artvervir	c_Revtraviricetes	o_Ortervirif	f_Retrovirif	g_Gammaretrovirus	s_Chick syncytial virus
3565	11867	k_Viruses	p_Artvervir	c_Revtraviricetes	o_Ortervirif	f_Retrovirif	g_Alpharetrovirus	s_Avian myelocytomatosis virus
3275	10261	k_Viruses	p_Nucleoc	c_Pokkesviricetes	o_Chitovirif	f_Poxviridag	g_Avipoxvirus	s_Fowlpox virus
1612	153135	k_Viruses	p_Artvervir	c_Revtraviricetes	o_Ortervirif	f_Retrovirif	g_Gammaretrovirus	s_
766	2844036	k_Viruses	p_Uroviric	c_Caudoviricetes	o_	f_Strabovirif	g_Pseudotevenvirus	s_lee
605	558016	k_Viruses	p_	c_Naldaviricetes	o_Lefavirif	f_Baculovif	g_Alphabaculovirus	s_
511	3051997	k_Viruses	p_	c_Naldaviricetes	o_Lefavirif	f_Baculovif	g_Betabaculovirus	s_chofumiferanae
394	11958	k_Viruses	p_Artvervir	c_Revtraviricetes	o_Ortervirif	f_Retrovirif	g_Alpharetrovirus	s_Avian carcinoma virus
334	755272	k_Viruses	p_Nucleoc	c_Megaviricetes	o_Algavirif	f_Phycodnif	g_Prasinivirus	s_Micromonas pusilla virus 12T
265	985782	k_Viruses	p_Nucleoc	c_Megaviricetes	o_Imitervif	f_Mimivirif	g_	s_Moumouvirus
241	2955291	k_Viruses	p_Negarnac	c_Insthoviricetes	o_Articulaf	f_Orthomyg	g_Alphainfluenzavirus	s_influenzae
240	2956167	k_Viruses	p_Lenarvir	c_Miaviricetes	o_Ourlivirif	f_Botourmg	g_Magoulivirus	s_niplasmoparae
204	3060017	k_Viruses	p_Nucleoc	c_Megaviricetes	o_Imitervif	f_Mimivirif	g_Fadolivirus	s_algeromassiliense
169	3047794	k_Viruses	p_Nucleoc	c_Megaviricetes	o_Pimascof	f_Ascovirif	g_Toursvirus	s_dptv1a
157	11866	k_Viruses	p_Artvervir	c_Revtraviricetes	o_Ortervirif	f_Retrovirif	g_Alpharetrovirus	s_Avian myeloblastosis virus
147	2956482	k_Viruses	p_Negarnac	c_Monjiviricetes	o_Mononef	f_Rhabdovg	g_Sunrhavirus	s_matariya
141	2107709	k_Viruses	p_	c_	o_	f_	g_Pandoravirus	s_quercus
127	3048337	k_Viruses	p_	c_Naldaviricetes	o_Lefavirif	f_Baculovif	g_Betabaculovirus	s_spliturae
124	1982251	k_Viruses	p_Uroviric	c_Caudoviricetes	o_	f_	g_Pahexavirus	s_
123	2956166	k_Viruses	p_Lenarvir	c_Miaviricetes	o_Ourlivirif	f_Botourmg	g_Magoulivirus	s_miplasmoparae
112	2588487	k_Viruses	p_Uroviric	c_Caudoviricetes	o_	f_Kyanovirif	g_	s_Synechococcus phage S-SCSM1
111	2126985	k_Viruses	p_Nucleoc	c_Megaviricetes	o_Imitervif	f_Mimivirif	g_Tupanvirus	s_soda lake
103	2560086	k_Viruses	p_Uroviric	c_Caudoviricetes	o_	f_	g_Alcyoneusvirus	s_
97	1985183	k_Viruses	p_Uroviric	c_Caudoviricetes	o_	f_Herellevg	g_Tsarbombavirus	s_tsarbomba
95	1094892	k_Viruses	p_Nucleoc	c_Megaviricetes	o_Imitervif	f_Mimivirif	g_Mimivirus	s_Megavirus chiliensis
91	2023057	k_Viruses	p_Nucleoc	c_Megaviricetes	o_Algavirif	f_Phycodnif	g_	s_Orpheovirus IHUM11002

BLAST of the CSV polymerase gene



Job Title	ref NC_043196.1
RID	4VB83EH3016 Search expires on 05-23 16:29 pm Download All ▾
Program	BLASTN ? Citation ▾
Database	nt See details ▾
Query ID	NC_043196.1
Description	Chick syncytial virus polymerase (pol) gene, partial cds
Molecule type	nucleic acid
Query Length	1248
Other reports	Distance tree of results MSA viewer ?

Filter Results

Organism *only top 20 will appear* exclude

Type common name, binomial, taxid or group name

[+ Add organism](#)

Percent Identity to **E value** to **Query Coverage** to

[Filter](#) [Reset](#)

- Descriptions**
- Graphic Summary
- Alignments
- Taxonomy

Sequences producing significant alignments

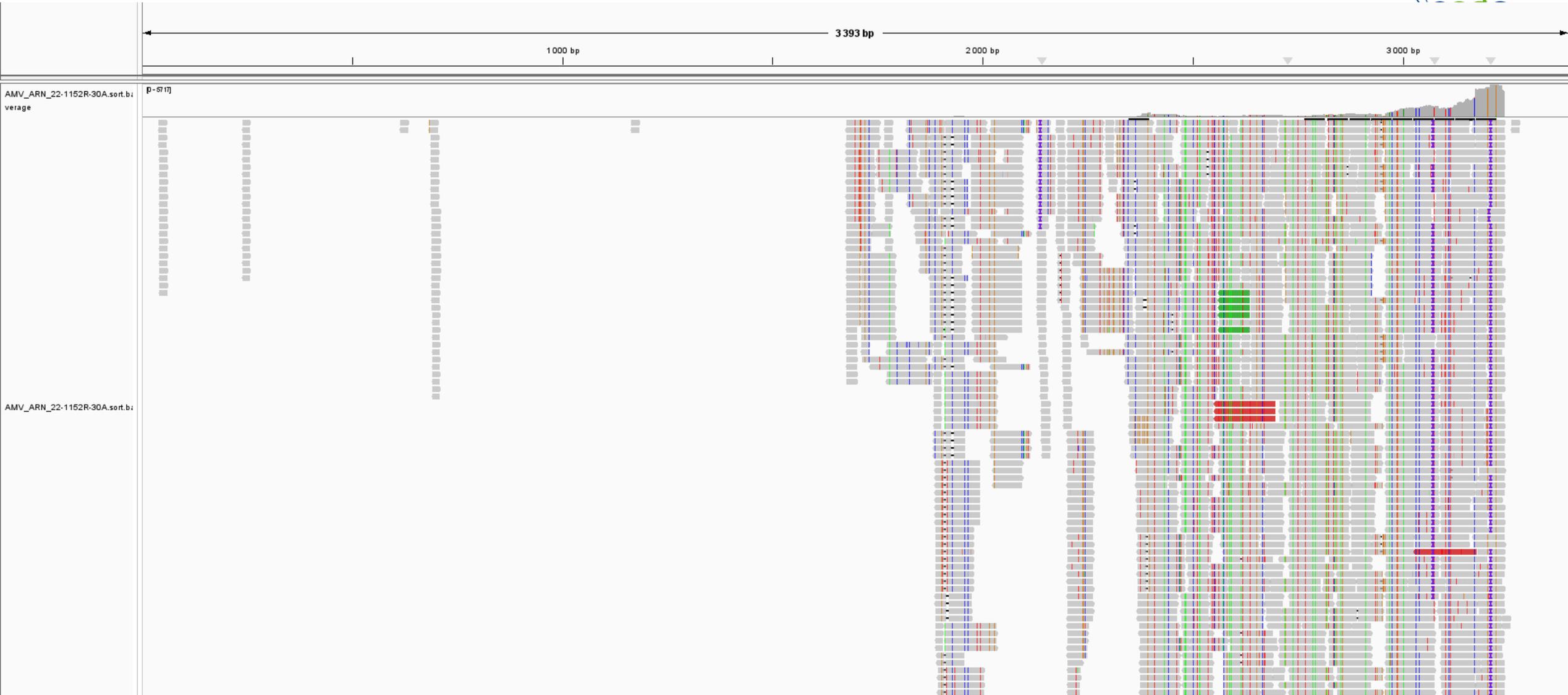
Download ▾ Select columns ▾ Show [?](#)

select all *88 sequences selected*

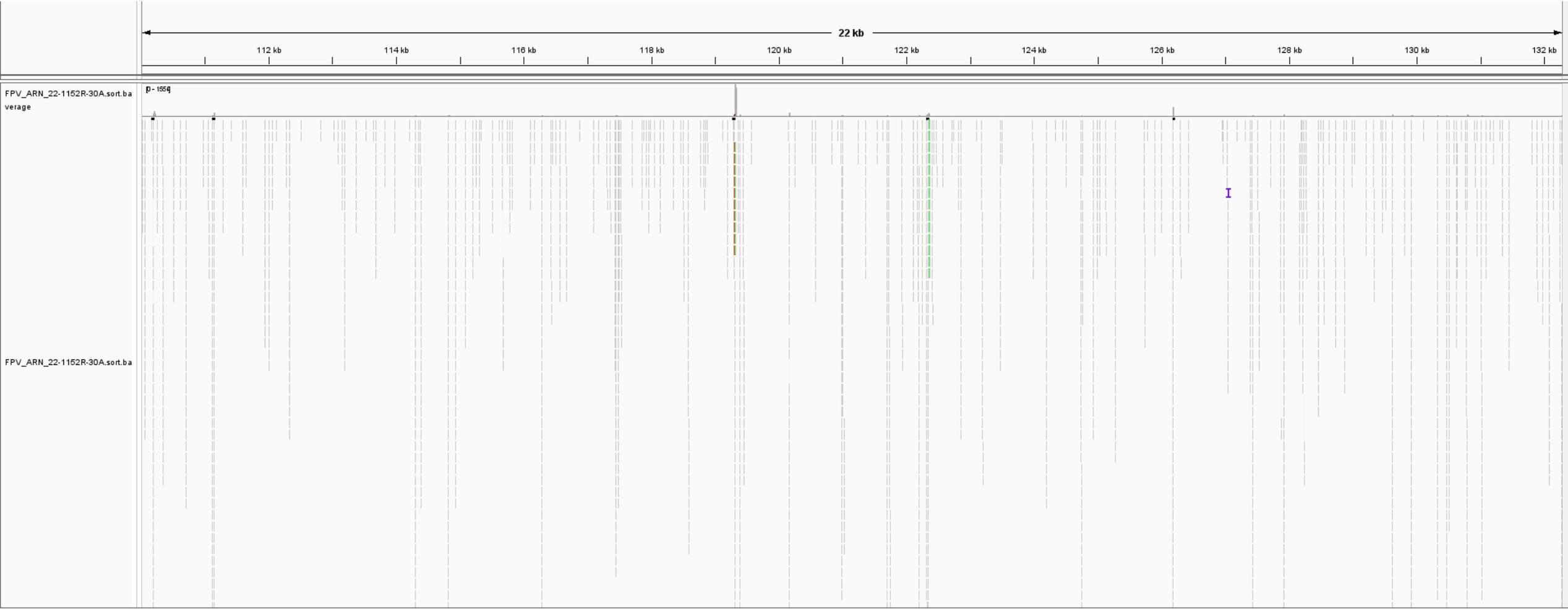
[GenBank](#) [Graphics](#) [Distance tree of results](#) [MSA Viewer](#)

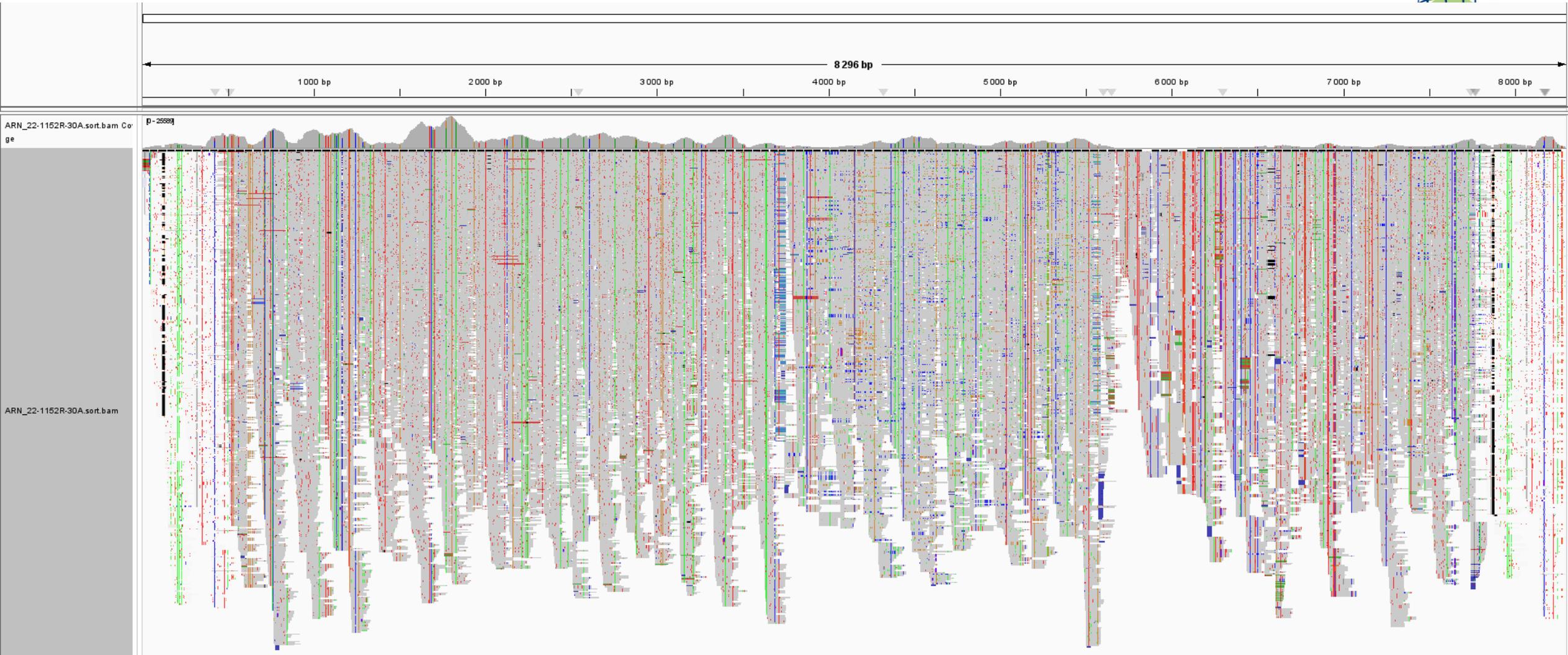
	Description	Scientific Name	Max Score	Total Score	Query Cover	E value	Per. Ident	Acc. Len	Accession
<input checked="" type="checkbox"/>	Chick syncytial virus polymerase (pol) gene, partial cds	Chick syncytial ...	2305	2305	100%	0.0	100.00%	1248	NC_043196.1
<input checked="" type="checkbox"/>	Reticuloendotheliosis virus isolate TX-01 polymerase (pol) gene, partial cds	Reticuloendothe...	2278	2278	100%	0.0	99.60%	1253	DQ237906.1
<input checked="" type="checkbox"/>	Reticuloendotheliosis virus isolate REV-TH/CBI/2013/CU-1, complete genome	Reticuloendothe...	2272	2272	100%	0.0	99.52%	8284	MF631845.1
<input checked="" type="checkbox"/>	Fowlpox virus isolate FWPV-SD15-670_2	Fowlpox virus	2272	2272	100%	0.0	99.52%	295938	MH734528.1

Avian myelocytomatosis virus



Fowlpox virus





Integrative genomics viewer

James T Robinson [✉](#), Helga Thorvaldsdóttir, Wendy Winckler, Mitchell Guttman, Eric S Lander, Gad Getz & Jill P Mesirov [✉](#)

Shotgun metagenomics, Example of Bat CoV sequencing

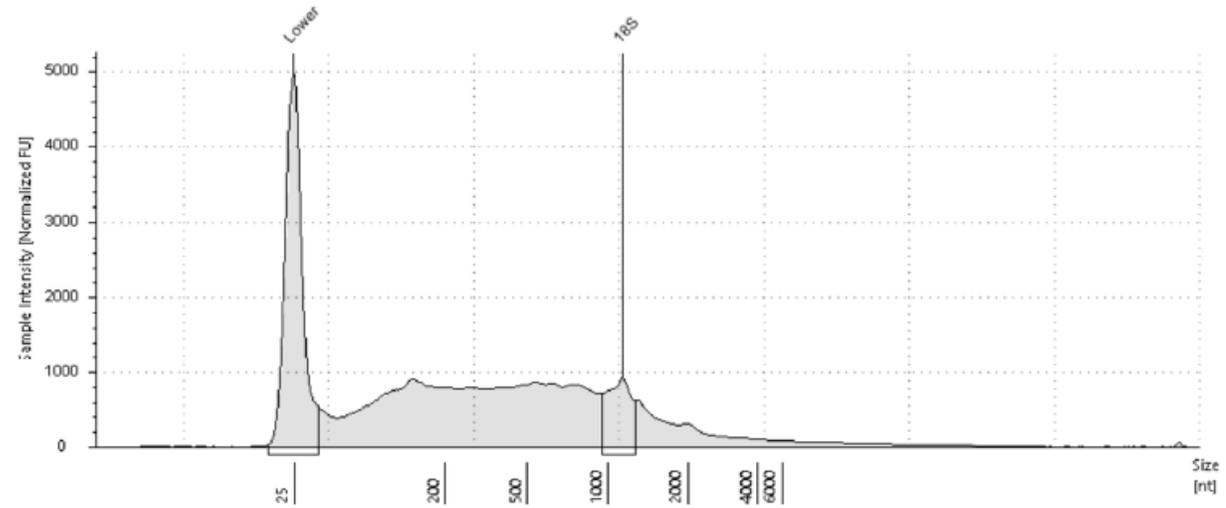
Metagenomics analysis of bat droppings



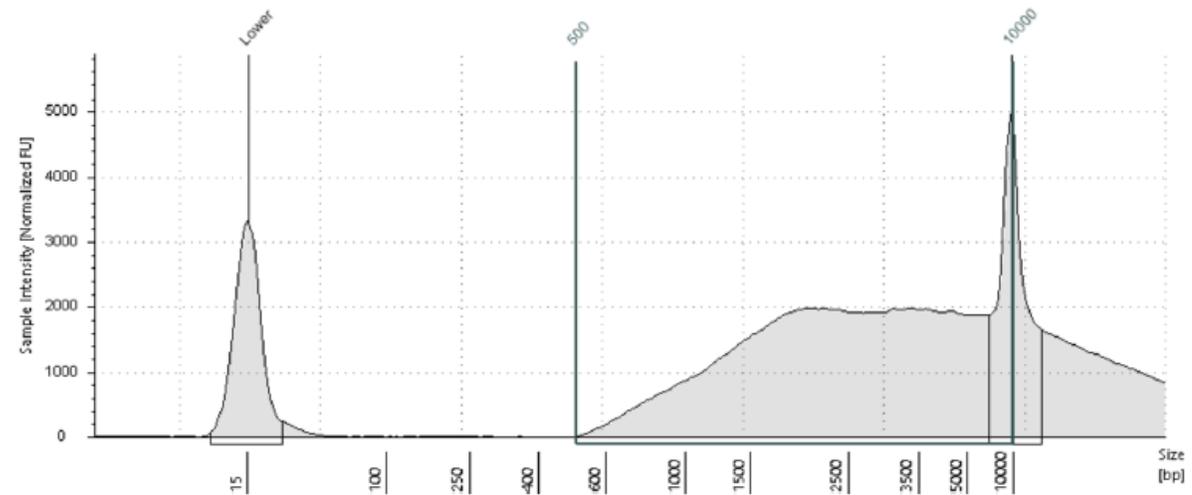
- **Sample type:** bat guano
- **Wetlab:**
 - Nucleic acids extraction (magnetic beads, Mag Fast protocol)
 - Random amplification of RNA (SMART9N protocol - Claro *et al.* 2023)
 - Oxford Nanopore Technologies sequencing
- **In silico:**
 - Taxonomic sequence classification (Kraken2 - Wood *et al.* 2019)
 - Sequence alignment (Minimap2 - Li, H. 2018)

Examples of TapeStation results

Total RNA



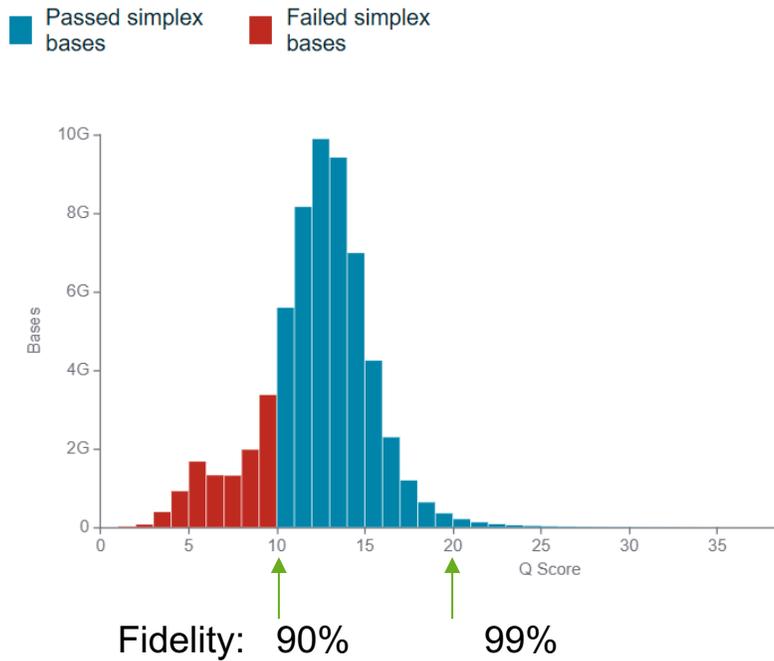
ds cDNA
(SMART9N amplification)



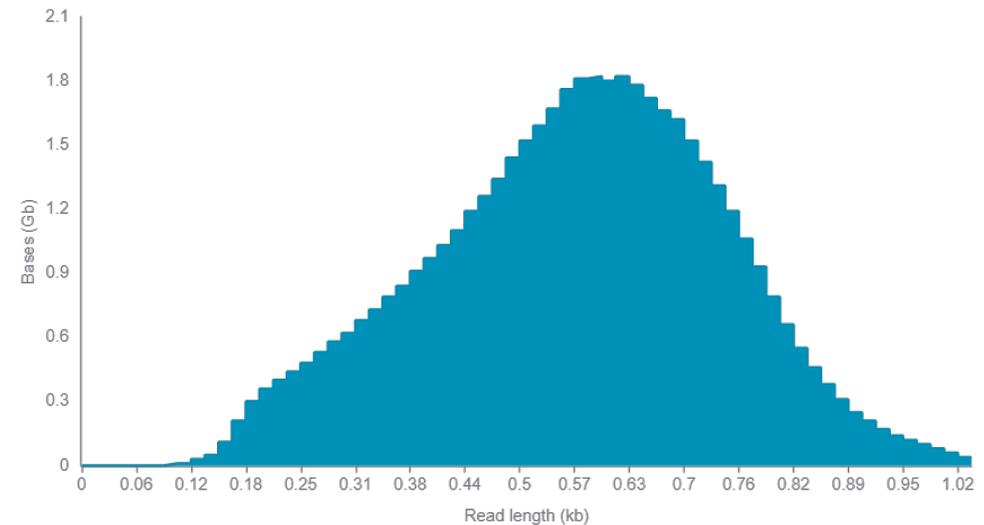
Results

- 21 bat samples sequenced using PromethION flowcells on P2Solo device.
- 125 millions reads (64 Gigabases) with a minimum Qscore of 10.

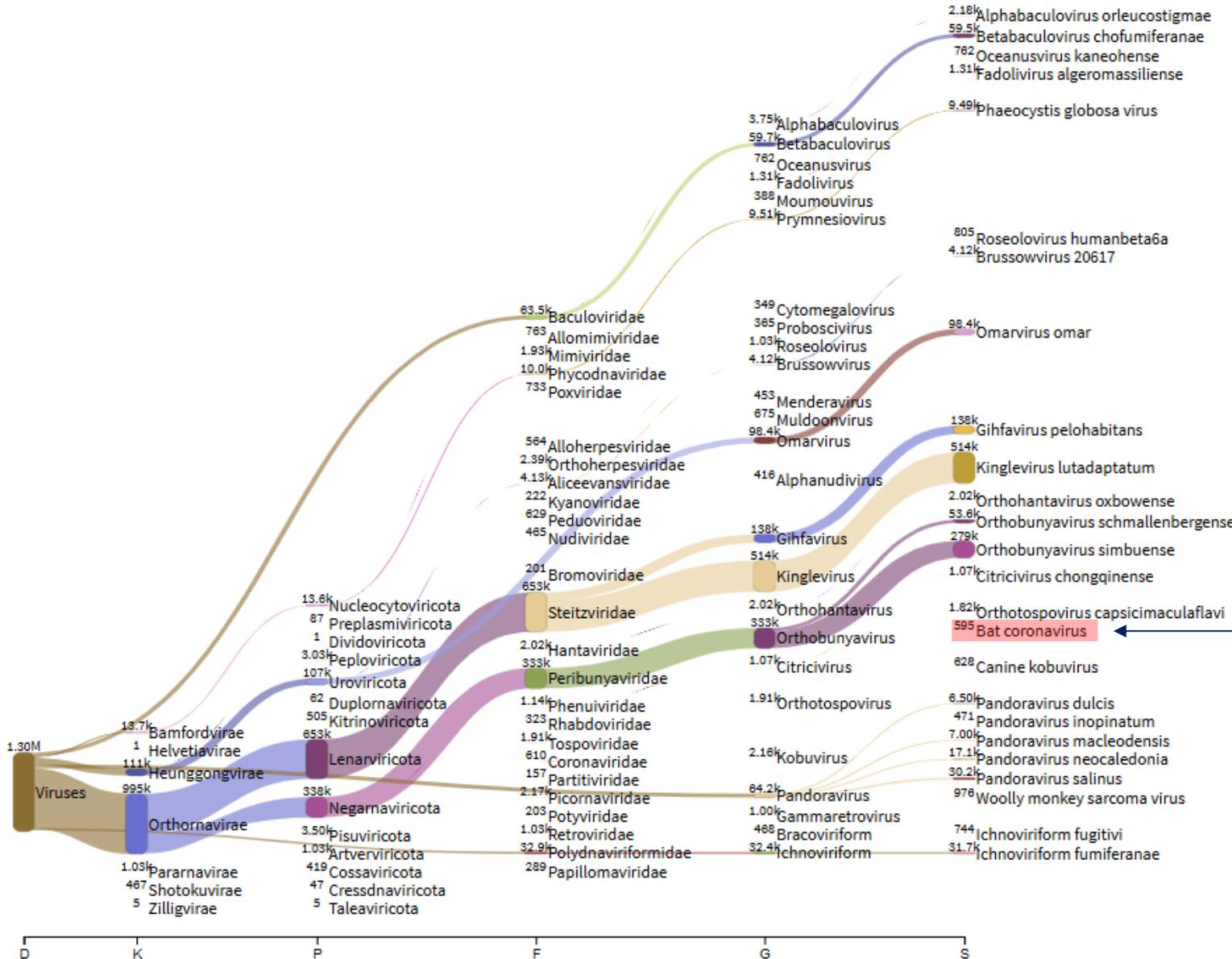
Q Score histogram



- Size distribution, up to 1Kb



- Taxonomic sequence classification (Kraken2, Wood et al. 2019)



Pavian: interactive analysis of metagenomics data for microbiome studies and pathogen identification FREE

Florian P Breitwieser ✉, Steven L Salzberg

- BLAST analysis to confirm taxonomic affiliation



Results for

Program **BLASTN** [Citation](#)

Database **core_nt** [See details](#)

Query ID **lcl|Query_3434576**

Description **ba257400-4622-47d0-82a8-a119e38d987c_runid=e009462 ...**

Molecule type **dna**

Query Length 1568

Other reports [Distance tree of results](#) [MSA viewer](#)

Organism *only top 20 will appear* exclude

[+ Add organism](#)

Percent Identity to

E value to

Query Coverage to

Filter **Reset**

- Descriptions**
- Graphic Summary
- Alignments
- Taxonomy

Sequences producing significant alignments Download Select columns Show

select all *100 sequences selected* [GenBank](#) [Graphics](#) [Distance tree of results](#) [MSA Viewer](#)

	Description	Scientific Name	Max Score	Total Score	Query Cover	E value	Per. Ident	Acc. Len	Accession
<input checked="" type="checkbox"/>	Bat coronavirus isolate CMR705-P13 ORF1a, ORF1b, Spike protein, ORF3, Membrane protein, capsid, hypotheti...	Bat coronavirus	2776	2776	100%	0.0	98.60%	28958	MG693172.1
<input checked="" type="checkbox"/>	Bat coronavirus, complete genome	Bat coronavirus	2771	2771	100%	0.0	98.54%	28975	NC_048212.1
<input checked="" type="checkbox"/>	Eidolon bat coronavirus/Kenya/KY24/2006 polyprotein (ORF1ab) gene, partial cds; and spike protein (S), ORF3 p...	Eidolon bat coro...	2743	2743	100%	0.0	98.22%	16186	HQ728482.1
<input checked="" type="checkbox"/>	Bat coronavirus isolate E.helvum, Nigeria_37_2022, partial genome	Bat coronavirus	2687	2687	100%	0.0	97.58%	28975	PP860750.1
<input checked="" type="checkbox"/>	Eidolon bat coronavirus isolate 5B/Kenya/BAT1994/2015, complete genome	Eidolon bat coro...	2643	2643	100%	0.0	97.07%	29061	PP273180.1
<input checked="" type="checkbox"/>	Eidolon bat coronavirus isolate 5A/Kenya/BAT594/2015, complete genome	Eidolon bat coro...	2632	2632	100%	0.0	96.95%	29085	PP273179.1
<input checked="" type="checkbox"/>	Bat coronavirus isolate CMR891-892 ORF1a gene, partial cds; and ORF1b, Spike protein, ORF3, Membrane prot...	Bat coronavirus	2610	2610	100%	0.0	96.69%	28810	MG693171.1

- Sequence alignment vs reference genome

BatCoV reference genome
(28 Kb)



In summary

Oxford Nanopore Technologies sequencing:

- Can be used on the field
- Allows outbreak tracking
- Allows shotgun metagenomics studies

Acknowledgements

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Team VIRAL

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